

COVID vaccination and post-infection cancer signals: Evaluating patterns and potential biological mechanisms

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Keywords: COVID; vaccine; cancer; infection; lymphoma; leukemia; sarcoma; carcinoma

Received: November 26, 2025 **Accepted:** December 26, 2025 **Published:** January 03, 2026

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ABSTRACT

A growing number of peer-reviewed publications have reported diverse cancer types appearing in temporal association with COVID-19 vaccination or infection. To characterize the nature and scope of these reports, a systematic literature search from January 2020 to October 2025 was conducted based on specified eligibility criteria. A total of 69 publications met inclusion criteria: 66 article-level reports describing 333 patients across 27 countries, 2 retrospective population-level investigations (Italy: ~300,000 cohort, and Korea: ~8.4 million cohort) quantified cancer incidence and mortality trends among vaccinated populations, and one longitudinal analysis of ~1.3 million US military service members spanning the pre-pandemic through post-pandemic periods. Most of the studies documented hematologic malignancies (non-Hodgkin's lymphomas, cutaneous lymphomas, leukemias), solid tumors (breast, lung, melanoma, sarcoma, pancreatic cancer, and glioblastoma), and virus-associated cancers (Kaposi and Merkel cell carcinoma). Across reports, several recurrent themes emerged: (1) unusually rapid progression, recurrence, or reactivation of preexisting indolent or controlled disease, (2) atypical or localized histopathologic findings, including involvement of vaccine injection sites or regional lymph nodes, and (3) proposed immunologic links between acute infection or vaccination and tumor dormancy, immune escape, or microenvironmental shifts. The predominance of case-level observations and early population-level data demonstrates an early phase of potential safety-signal detection. These findings underscore the need for rigorous epidemiologic, longitudinal, clinical, histopathological, forensic, and mechanistic studies to assess whether and under what conditions COVID-19 vaccination or infection may be linked with cancer.

INTRODUCTION

The COVID-19 pandemic and the widespread deployment of novel mRNA- and viral-vector based

vaccines have reshaped the landscape of human immunology [1–4]. Never has such a large proportion of the global population been exposed simultaneously to nucleic acid-based immunogens, lipid nanoparticle (LNP)

delivery systems, and repeated booster regimens over a relatively short period. The unprecedented scale that was marshaled in response to the COVID-19 pandemic has generated and continues to generate extensive clinical, molecular, and epidemiologic data, revealing biological responses that extend beyond traditional vaccine-induced immune activation and responses. These include a spectrum of post-infection and post-vaccination neurological, autoimmune, and inflammatory syndromes, including myocarditis, immune-mediated neuropathies, autoimmune cytopenias, systemic inflammatory responses [5–7], as well as temporal co-occurrence with cancer diagnoses, recurrences, or unexpectedly rapid disease trajectories [8–11]. These events have prompted extensive clinical investigation and underscore the capacity of vaccine-induced immune activation to perturb immune homeostasis in susceptible individuals. Importantly, many of these conditions are characterized by cytokine dysregulation, altered innate and adaptive immune signaling, and tissue-specific inflammatory responses; pathways that are also implicated in tumor initiation, progression, and immune surveillance. The present review focuses specifically on cancer-related observations within this broader context of post-vaccination immune perturbation.

After nearly six years since the pandemic was recognized in early 2020, the current world's literature addressing COVID-19 infection or vaccination and cancer remains sparse, heterogeneous, and largely limited to case reports and small case series, insufficient to support definitive conclusions regarding causation or quantification of risk. Package inserts for COVID19 vaccines posted by the Food and Drug Administration (FDA) [12–15] specifically state that they have not been evaluated for carcinogenicity or genotoxicity, nor have they been studied after multiple vaccine doses and boosters or in combination with subsequent SARS-CoV-2 infection.

During the COVID pandemic, it was predicted that cancer rates would rise during and after COVID due to reduced screening and reduced access to treatment during the pandemic. However, rates of cancer among younger individuals for example with early onset colon cancer have been rising for two decades [16, 17]. Rates of cholangiocarcinoma and endometrial cancer have been rising as well. Cancer deaths exceeded 600,000 in US for 1st time in 2024 and in 2025 are predicted to rise as well [18]. As of the writing of this review, there are no published population studies in the US with mortality or cancer incidence follow-up beyond 42 days for outcomes after Covid infection versus no Covid infection or Covid vaccinated versus not Covid vaccinated. This is in part due to lack of good quality databases that would have such information. There is a National Cancer Institute (NCI)-funded Covid and Cancer Consortium (CCC) but it has not published on this topic specifically.

Against the backdrop of limited clinical evidence and incomplete preclinical toxicology, a recent study reported that SARS-CoV-2 mRNA vaccines may actually sensitize tumors to immune checkpoint blockade [19] prompting broad interpretation that COVID-19 mRNA vaccination may actually potentiate antitumor responses in patients with melanoma or non-small cell lung cancer (NSCLC) undergoing immune checkpoint inhibition. Moreover, in the analysis, mRNA vaccination was associated with increased Type I interferon signaling and elevated tumor PD-L1 expression. However, PD-L1 upregulation in the absence of checkpoint inhibitor therapy is generally associated with enhanced tumor immune evasion and resistance to T-cell-mediated cytotoxicity, raising questions about the biological interpretation of these findings. Although interferon-based therapies have established clinical utility in melanoma, the study did not provide comparative analyses between interferon treatment and the combination of mRNA vaccination with checkpoint blockade. Furthermore, the study did not address key limitations, alternative mechanistic explanations, or the broader clinical context necessary to fully interpret the reported effects.

This absence of evaluation of COVID19 vaccines for carcinogenicity or genotoxicity motivated a systematic review and synthesis of the available evidence from 2020–2025 concerning COVID-19 vaccination, SARS-CoV-2 infection, and cancer. Specifically, we sought to (i) categorize malignancies reported in temporal proximity to vaccination or infection, (ii) evaluate temporal and clinical patterns across tumor types for relevant signals among patients exposed to the COVID vaccines, and (iii) outline plausible immunologic and molecular mechanisms that could underlie these phenomena.

Across the published literature, we identified reports involving hematologic malignancies, including lymphomas and leukemias, solid tumors such as breast, lung, pancreatic, and glial cancers, virus-associated malignancies including Kaposi sarcoma and Merkel cell carcinoma, and rare entities such as sarcomas, melanomas, and adenoid cystic carcinomas. While the number of studies or their temporal association does not establish causation, understanding whether these associations represent coincidence, immune dysregulation, or a broader biologic effect linking infection, vaccination, and cancer development is now of pressing importance.

Importantly, regarding reported adverse events and potential risks, awareness of what has occurred, even if ultimately this proves to be extremely rare, is a necessary component of informed consent at a time when there is no longer a public health emergency from COVID-19. Cancer risk is likely based on heterogeneity among individuals, the impact of genetics, environment, and interacting social determinants of health that varies among individuals and this is an area where this article

could form a foundation for future studies to refine individualized risk. As such, the goal of this article is to systematically synthesize and contextualize findings from the published literature regarding malignancies temporally associated with COVID-19 vaccination or SARS-CoV-2 infection, without attempting to estimate risk, establish causality, or inform individual clinical or vaccination decisions.

RESULTS

This scoping review, covering the period of January 2020 until April 2025, was not designed to estimate cancer risk or incidence, nor to draw causal inferences, but rather to systematically assemble, categorize, and contextualize published reports of malignancies temporally associated with COVID-19 vaccination or SARS-CoV-2 infection. It identified 69 publications [8, 20–87] describing malignancies or malignant progression in temporal association with COVID-19 vaccination or SARS-CoV-2 infection, encompassing a total of 333 patients (excluding population-level studies [8, 20]). In addition, one population-level publication which offered a longitudinal assessment of cancer incidence across the pandemic and immediate post-pandemic period was identified [85]. Among the 69 studies, most reports were single-patient case reports or small series (55/69, 81%), with a small number of systematic or narrative reviews (3/69, 4.5%), mechanistic/experimental studies (2/69, 3%), and larger case series, multicenter, or database-level analyses (8/69, 12%) (Table 1). Consistent with an early signal-detection phase, the underlying evidence base is therefore heavily weighted toward documenting occurrences of potentially adverse events and hypothesis-generating case-level observations rather than population-based epidemiologic studies.

Geographic distribution

Reports originated from a wide range of countries spanning Asia, Europe, the Middle East, Africa, and North and South America. The countries with the highest number of publications were Japan ($n = 11$) and the United States ($n = 11$), followed by China ($n = 7$) and Italy ($n = 4$). Additional single-patient cases or small series were identified from Spain, South Korea, Saudi Arabia, India, Nigeria, Brazil, Turkey, Singapore, Lebanon, Egypt, Bulgaria, Taiwan, Ukraine, Iran, Russia, Greece, Austria, Germany, Poland/Ukraine, as well as multi-institutional or international collaborations. This broad geographic distribution indicates that the reported temporal associations between COVID-19 vaccination or infection and oncologic events are not confined to a particular region or healthcare system but have been observed across diverse clinical settings and diagnostic infrastructures around the globe.

Exposure types: Vaccination versus infection

Most publications identified in the search focused on oncologic events occurring after COVID-19 vaccination (56/69; 89%), with the remainder describing associations following SARS-CoV-2 infection (5/69; 7%), and SARS-CoV-2 infection with prior vaccination (7/69; 10%). One publication (1/69; 1%) did not explicitly specify whether the reported oncologic event followed vaccination, SARS-CoV-2 infection, or a combination of both exposures. These included case reports and mechanistic studies evaluating post-infectious tumor behavior, immune perturbation, or disease acceleration along with SARS-CoV-2 infection but in the absence of vaccination or associated with a SARS-CoV2 infection but with prior vaccination or boosting. The predominance of vaccination-associated case reports may reflect reporting patterns rather than comparative biological risk, and the available data lack sufficient individual-level detail to determine whether or how oncologic responses differ between infection or vaccination.

Across the published literature, reported vaccine formulations and exposure types were heterogeneous but could be grouped into broad platform categories (Figure 1). Among vaccine-related reports, the majority involved mRNA vaccines, with approximately 56% following the Pfizer-BioNTech vaccine (BNT162b2) and 25% following the Moderna vaccine (mRNA-1273). An additional 5% involved patients who had received both Pfizer and Moderna products across different doses. Adenovirus vector vaccines represented the next largest category, including AstraZeneca (ChAdOx1/Covishield) (5.8%), Johnson & Johnson (Ad26.COV2.S) (2.9%) and the Russian, Sputnik-V (1.4%). Inactivated vaccines (e.g., Sinopharm BBIBP-CorV, CoronaVac, or other formulations) and studies in which the specific vaccine type was not reported were least represented (2.6% and 1.1%, respectively). This distribution indicates that the published literature is heavily weighted toward mRNA vaccine platforms, particularly Pfizer-BioNTech and Moderna, which together account for the vast majority of vaccine-associated reports. This pattern closely mirrors global vaccination practices where mRNA vaccines were most widely deployed. The relatively smaller representation of adenoviral vector vaccines and inactivated platforms likely reflects both their more limited use in certain regions and differential reporting practices, rather than a comparative assessment of biological risk.

Cancer types and clinical spectrum

Approximately 43% (30/69) of publications reported lymphoid malignancies, encompassing both lymphomas and leukemias (Figure 2 and Table 2). These included a wide spectrum of lymphoid neoplasms such as diffuse large B-cell lymphoma (DLBCL), various T-cell lymphomas (e.g., angioimmunoblastic T-cell lymphoma, subcutaneous

Table 1: Summary of reports linking COVID-19 vaccination or infection to cancer

Study type	N	% of Total (N = 69)	Comments
Case reports	50	72%	Dominant study type; mostly single-patient descriptions
Case series	5	7%	Typically 2-several patients
Systematic/narrative reviews	3	4%	Summaries or literature syntheses
Cohort/retrospective/observational population studies	8	12%	Larger-scale data (e.g., population cohort, single center cohort)
Mechanistic/translational studies (tissue, organoids, mouse)	3	4%	Experimental or preclinical mechanistic work

pannicketitis-like T-cell lymphoma), chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL), and cutaneous T-cell lymphomas (CTCL). Several reports emphasized unexpectedly rapid progression, atypical presentations, or unusually aggressive courses of disease.

Solid tumors accounted for 41% of publications (28/69) and represented a diverse group of malignancies, including melanoma, breast cancer, lung cancer, glioblastoma and other glial tumors, sarcomas, and various organ-specific carcinomas, such as pancreatic cancer (Figures 2 and 3). In multiple reports, the authors

described unusually rapid onset, short-latency recurrence, or aggressive clinical progression for tumor types such as pancreatic adenocarcinoma and glioblastoma; features that are atypical for these cancers highlighted as notable temporal observations.

A subset of reports described tumor formation or recurrence at or near vaccine injection sites, the deltoid region, axilla, or draining lymphatic basins, including cases where axillary lymphadenopathy coincided with solid-tumor metastasis. Virus-associated malignancies such as Kaposi sarcoma, Merkel cell carcinoma, and EBV-

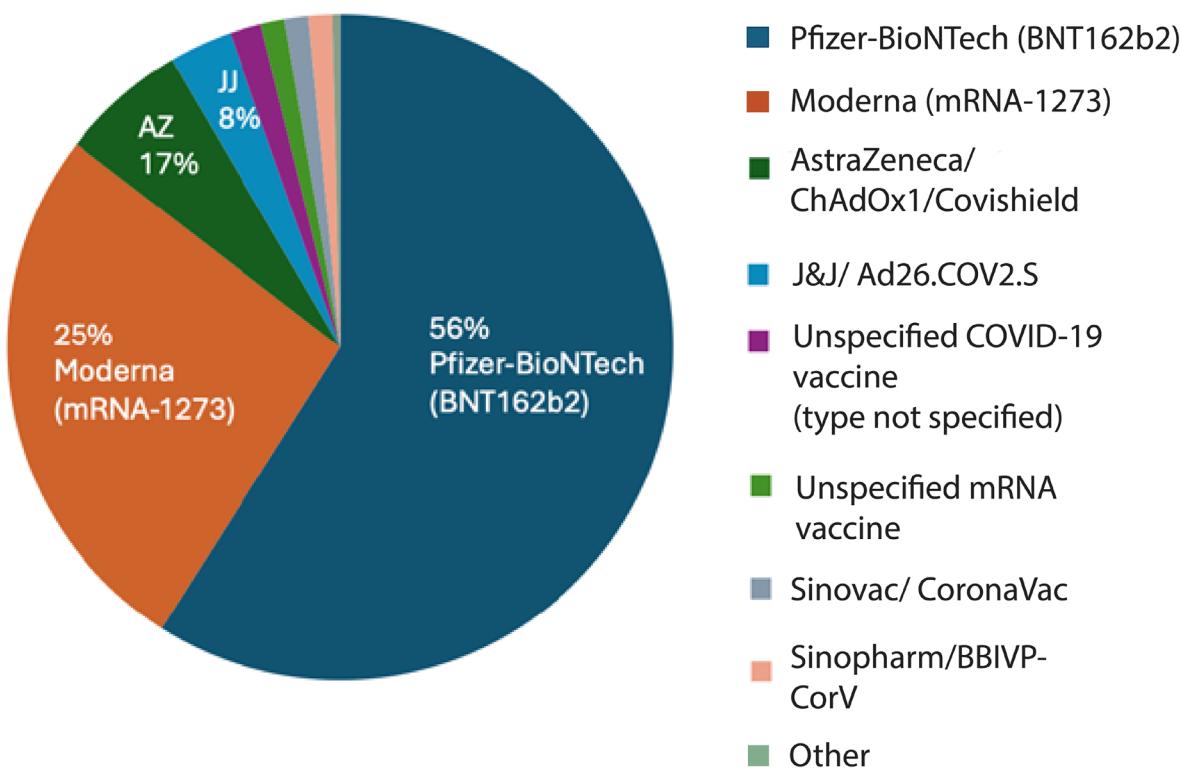


Figure 1: Distribution of reported malignancies by COVID-19 vaccine type. Distribution of vaccine formulations among vaccinated patients with reported cancer following COVID-19 immunization. Most cases involved Pfizer-BioNTech (BNT162b2; 56%) and Moderna (mRNA-1273; 25%) vaccines, followed by AstraZeneca/ChAdOx1 (Covishield; 17%) and Johnson & Johnson/Ad26.COV2.S (8%). A small fraction of reports involved Sinovac (CoronaVac), Sinopharm (BBIBP-CorV), or other inactivated vaccines, as well as unspecified mRNA or COVID-19 vaccine types. The predominance of mRNA vaccines reflects their widespread global use during the study period.

positive lymphomas were also identified across several reports. The remaining 16% of publications (11/69) were categorized as other or unspecified, which included mixed or indeterminate cases, non-malignant proliferations,

studies referencing “cancer”, “tumor”, or “malignancy” without definitive histopathologic classification, and population-level analyses in which tumor type was not explicitly delineated.

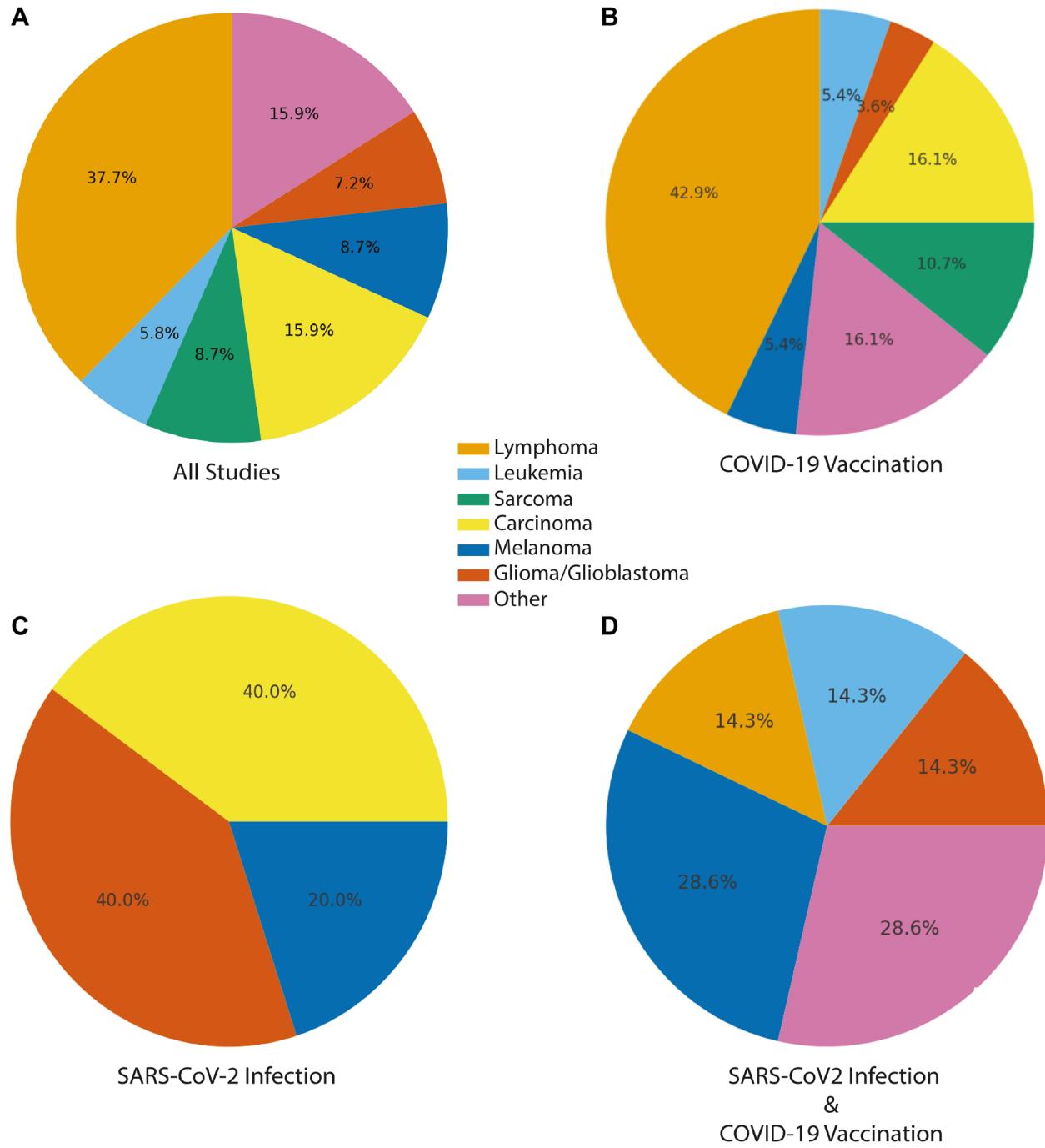


Figure 2: Distribution of post-vaccination and post-infection malignancies by tumor type. Distribution of reports with malignancy or tumor-like lesions temporally associated with COVID-19 vaccination, SARS-CoV-2 infection, or SARS-CoV-2 infection and vaccination. Pie charts depict the proportional representation of major cancer categories observed. (A) Across all studies. (B) COVID-19 vaccination, (C) SARS-CoV-2 infection, and (D) combined SARS-CoV-2 infection and COVID-19 vaccination. Cancer types were consolidated into seven high-level categories. Carcinoma includes: breast cancer, prostate cancer, colon cancer, pancreatic cancer, lung cancer, Merkel cell carcinoma, GI neoplasia/polyposis. Lymphoma also includes lymphoid neoplasms, cutaneous lymphoproliferative disorders, lymphoproliferative disorder. Other includes benign tumors, pseudotumors, mixed tumors, heart tumors, inflammatory and non-specific tumors (e.g., myofibroblastic).

Table 2: Clinicopathologic spectrum of lymphomas in post-vaccination reports

Lineage	Subtypes	Key features
<i>T-cell lymphomas</i>	CTCL, LyP, ALCL, AITL, SPTCL, TFH-type, PCGDTCL, T-ALL, T-cellNOS	Dominated by cutaneous and TFH-derived entities; several at injection sites; many indolent or self-resolving (CD30 ⁺).
<i>B-cell lymphomas</i>	DLBCL, Follicular, MZL, CLL	Primarily DLBCL; often nodal or axillary post-mRNA vaccine; typically de novo; most treated with R-CHOP.
<i>NK/NK-T-cell lymphomas</i>	ENKL (nasal-type), NK/T overlap	EBV ⁺ nasal lesions; one partial response to SMILE + radiation; suggest EBV reactivation.
<i>Mixed/Unspecified LPDs</i>	Large “unspecified/other” cohort from systematic review (Cui 2024) and PCLDs	Aggregate data without cell-lineage resolution; largely literature or registry series.

Specific examples of cancers and their association with COVID vaccination

Lymphoma

Cavanna et al. [26] reports the review of a series of eight patients who developed Non-Hodgkin's Lymphoma after COVID-19 vaccination (Table 3), including four males and four women. Five patients were vaccinated with the BNT162b2 vaccine (Pfizer), one with the ChAdOx1 nCOV-19 vaccine (AstraZeneca, Cambridge, UK), one with mRNA 1273/Spikevax (ModernaTX) and one patient with the recombinant replication-incompetent adenovirus type 26 (Ad26) viral-vector-based COVID-19 vaccine (Janssen Pharmaceuticals, Beerse, Belgium). One of the NHL cases presented with large right axillary adenopathy shortly after COVID-19 vaccination (Figure 3A).

Sekizawa et al. [28] describe a case of marginal zone B-Cell lymphoma in an 80-year-old Japanese woman who presented with a right temporal mass that appeared the morning after she was administered her first mRNA COVID-19 vaccination (BNT162b2) (Figure 3B). The mass gradually decreased in size but persisted over 6 weeks after her first vaccination (3 weeks after her second vaccination). At her first visit, ultrasound revealed the size of the mass to be 28.5 Å~ 5.7 mm, and computed tomography revealed multiple lymphadenopathies in the right parotid, submandibular, jugular, and supraclavicular regions. This case brings up the possibility that an initial mass may not be composed entirely of cancer cells and may have an element of a host response that may limit the progression depending on immune or other factors. In this case, the patient had marginal zone B-cell lymphoma after BNT162B2 COVID-19 vaccination.

Sarcoma

Bae et al. [21] reported the development of high grade sarcoma after the second dose of the Moderna vaccine. A 73-year-old female with a past medical history of hypertension, hyperlipidemia, and renal angiomyolipoma

status post resection in 2019 presented with worsening right upper arm swelling for the past two weeks. She noticed the swelling two to four days after receiving her second dose of the Moderna vaccine within 1 cm from the prior injection site. Physical examination was remarkable for a 6 cm, circular, mobile, soft mass present in the right upper arm. (Figure 3C). Li et al. [23] reported the development of classic cutaneous Kaposi's sarcoma in a 79-year-old male following the first dose of the ChAdOx1 nCov-19 vaccine, without prior SARS-CoV-2 infection or history of HIV infection. The patient developed multiple reddish-blue papules on his legs and feet, confirmed as KS through histopathology (Figure 3D). Treatment included radiotherapy and sequential chemotherapy with doxorubicin. The potential reactivation of latent HHV-8 by the vaccine is suggested through mechanisms involving the SARS-CoV-2 spike protein and adenovirus vector, which may induce immune responses and inflammatory pathways.

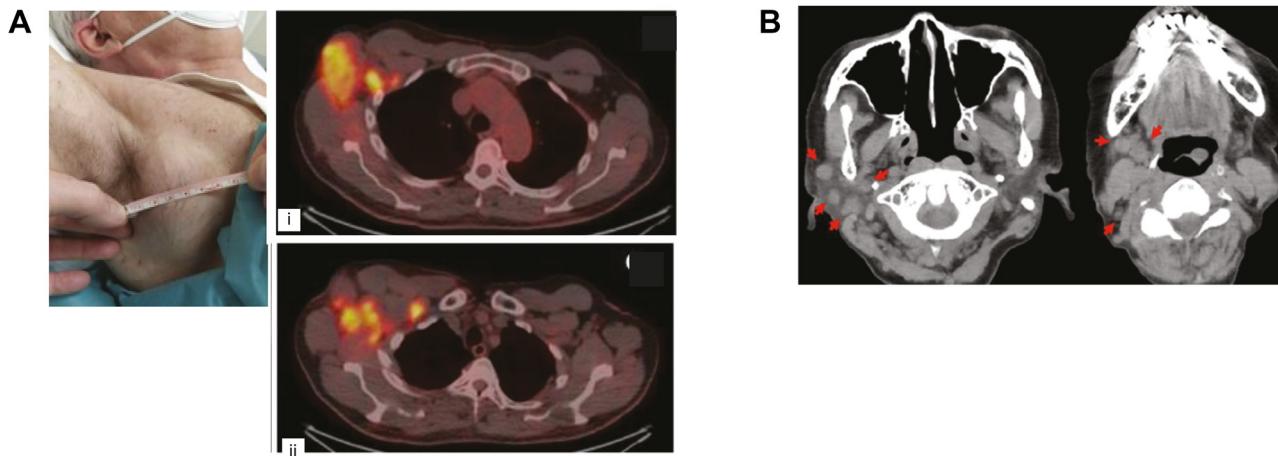
Carcinoma

Abue et al. [32] describe a case series of 96 patients with the diagnosis of pancreatic ductal adenocarcinoma (Figure 3E). Repeated COVID-19 booster vaccinations were associated with worse overall survival in the patients with pancreatic cancer. Analysis revealed that high levels of IgG4, induced by vaccination, correlate with a poor prognosis. Sano [36] described an 85-year-old woman who presented with an asymptomatic skin lesion in the right chest within one month immediately after the 6th dose of (Pfizer-BioNTech) vaccination. The patient had been diagnosed with right breast cancer two years prior and underwent partial mastectomy, hormone therapies, and was deemed to be in remission. The lesion was confirmed as a skin metastasis deemed to have developed through potential local recurrence at surgical margins (Figure 3F).

Melanoma

Wagle et al. [56] described a 49-year-old Indian male who developed rapidly progressive vision loss

LYMPHOMA



SARCOMA

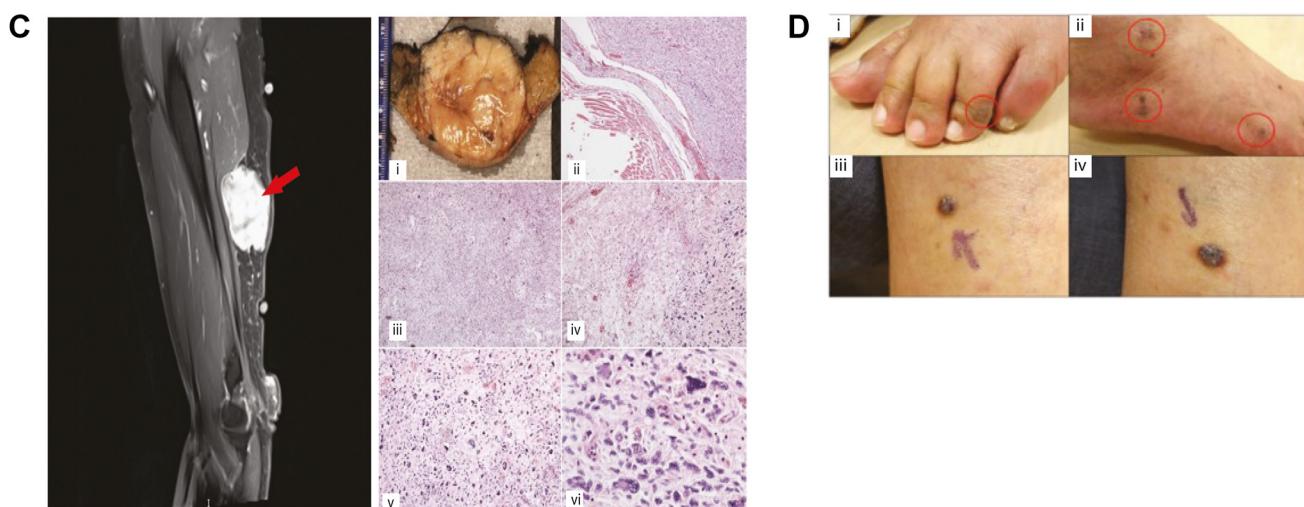
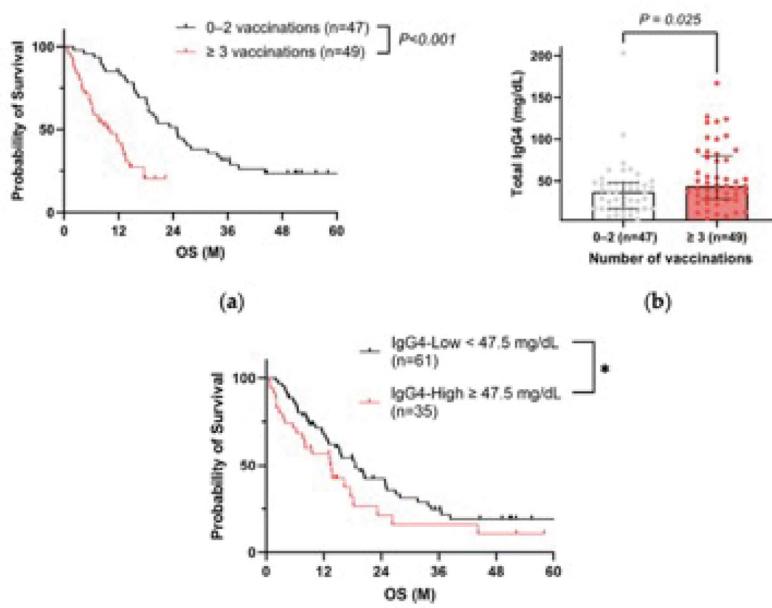


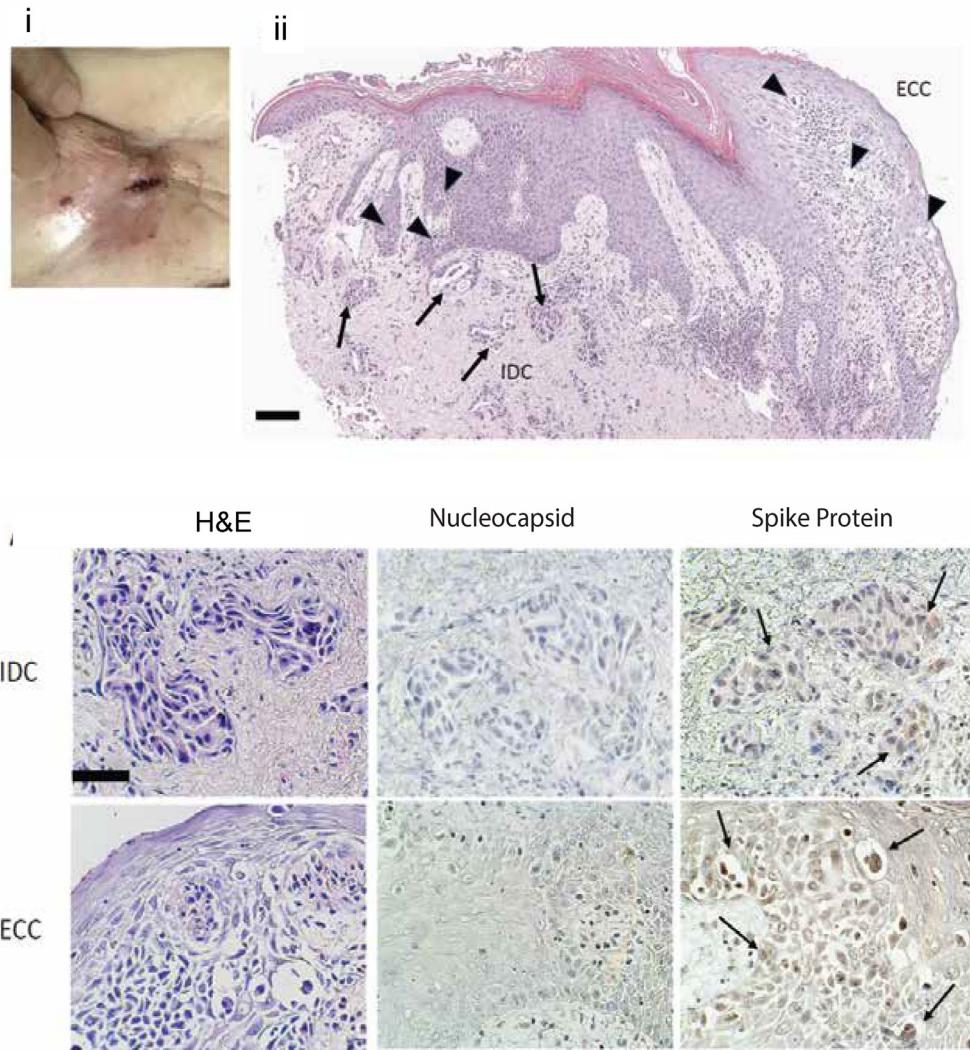
Figure 3: Representative examples of cancers reported in temporal association with COVID-19 vaccination. Figures were reproduced with permissions (Supplementary Table 1). Lymphoma: (A) Axillary adenopathy and i) 18-FDG-PET/CT at baseline in the right axillary adenopathy mass and ii) in multiple axillary adenopathies and subsequent NHL diagnosis following vaccination. Image reproduced from Cavanna et al., Medicina, 2023. © MDPI. (B) Temporal mass after her first BNT162b2 dose, with persistent lymphadenopathy on imaging. Axial computed tomography image shows (i, ii) submandibular and jugular regions. Image reproduced from Sekizawa et al., Front Med, 2022. © Frontiers. Sarcoma (C) High-grade sarcoma arising near injection site. A 6-cm right upper-arm mass after second Moderna dose, near the prior injection site; pathology confirmed high-grade sarcoma. Image adapted with permission from Bae et al., Cureus, 2023 © Springer Nature. (D) Classic cutaneous Kaposi's sarcoma adapted from Li et al. Front Med, 2022 © Frontiers. A 79-year-old man developed violaceous papules on the legs after the first ChAdOx1 vaccine dose; biopsy confirmed KS. Treatment included radiotherapy and doxorubicin. Clinical images of Kaposi sarcoma (i) with dark brown macules over the left foot, (ii) the right foot and larger reddish erythematous papules on his left calf (iii, iv). Carcinoma (E) In a 96-patient cohort, repeated booster vaccination correlated with poorer overall survival and elevated IgG4 level of pancreatic ductal adenocarcinoma. Kaplan-Meier analysis of 96 PC patients with known vaccination history and measured IgG4 levels, total IgG4 levels by number of vaccinations, and Kaplan-Meier analysis in PC patients by IgG4 levels. Image adapted with permission from Abue et al. Cancers 2025 © MDPI. (F) A case of metastatic breast carcinoma to the skin expressing SARS-CoV-2 spike protein. Histopathology of skin metasstatus along with IHC for nucleocapsid and Spike protein. Images adapted from Sano, S., J. Derm Sci, 2025. © Elsevier. Melanoma (G) Gross examination of specimen shows extensive intraocular hemorrhage involving both anterior and posterior chambers, accompanied by complete retinal detachment. H&E stained section shows

CARCINOMA

E

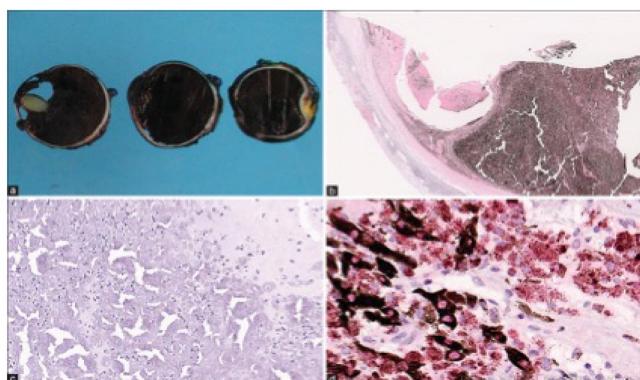


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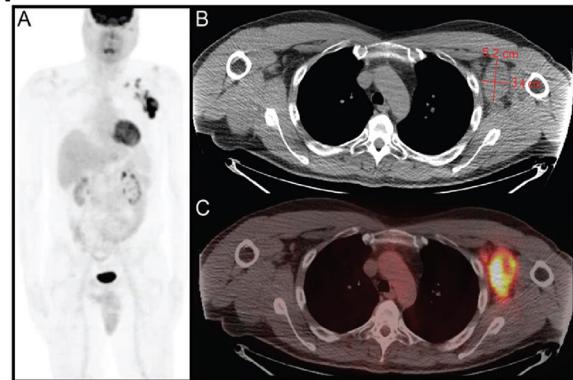


MELANOMA

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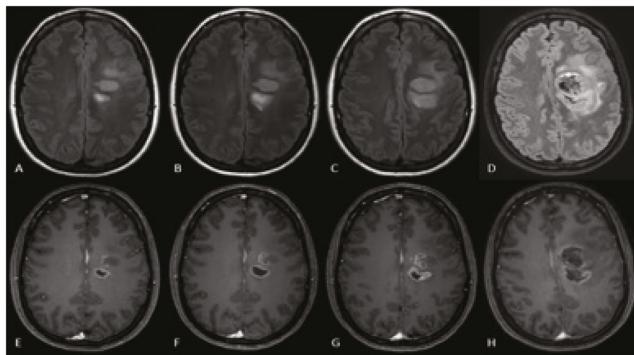


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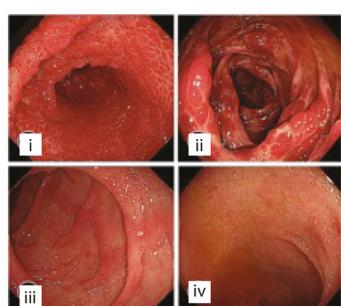
GLIOBLASTOMA

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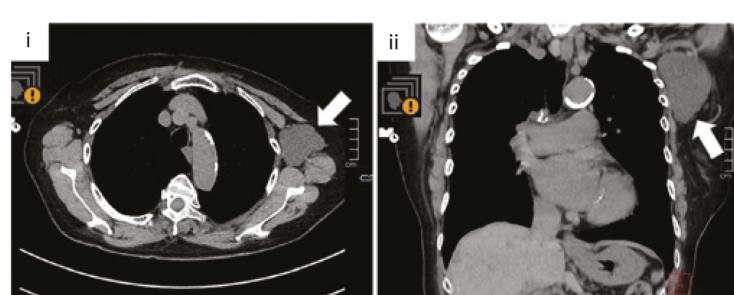


OTHER

J



K



severely degenerated, necrotic melanocytic lesion located with widespread necrosis within the melanocytic tumor. SOX10 IHC confirms melanocytic cells containing cytoplasmic melanin, interspersed among numerous SOX10-negative melanophages. Image adapted with permission from Wagle et al. Indian J Ophthalmol 2022 © Wolters Kluwer. (H) Maximum-intensity projection PET image shows markedly increased radiotracer uptake within the left axillary and supraclavicular lymph nodes. Representative axial CT and corresponding fused PET/CT images highlight the dominant nodal conglomerate. The patient had received a COVID-19 vaccination in the left upper arm within two months prior to imaging. Image adapted from Gullotti et al. Radiol Case Rep. 2022 © Elsevier. Glioblastoma (I) Two patients (ages 40 and 31) presented with new neurologic deficits and frontal-lobe masses shortly after mRNA vaccination. Image adapted from O'Sullivan et al. J of Neurology. 2021 © Elsevier. Other (J) Gastrointestinal polyposis identified following COVID-19 vaccination. Image adapted with permission from Kim et al. Clin Endosc 2024 © Korean Society of Gastrointestinal Endoscopy (K) Axillary lymphangioma in an 80-year-old woman three months after her second Pfizer-BioNTech dose; imaging showed a cystic lymphangioma. Image adapted with permission from Sasa et al. Surg Case Rep 2022 © Springer Nature.

Table 3: Summary of case series describing malignant lymphoma following mRNA COVID-19 vaccination

Case N	Gender/ Age (Year)	Time from Vaccination to Onset of Lymphoproliferative Disorder	Histopathological Examination	Type of COVID-19 Vaccine	Site and Diameter of Lymphadenopathy	Treatment of Lymphoma
1	M/67	1 day after 1 dose	DLBCL	BNT162b2	Left axilla 6.0 cm	Chemotherapy plus rituximab
2	F/80	2 days after 1 dose	DLBCL	BNT162b2	Left axilla 4.1 cm	Chemotherapy plus rituximab
3	F/58	7 days after 2 dose	DLBCL	BNT162b2	Left cervical area 4 cm	Radical surgery plus radiotherapy
4	M/53	3 days after 1 dose	Extranodal NK/T-cell lymphoma	BNT162b2	Erosive lesions upper lip up to 5 mm	Chemotherapy plus radiotherapy
5	M/51	7 days after 1 dose	EBV-positive DLBCL	ChAdox1 nCOV-19	Mediastinal mass 5 cm	Rituximab
6	F/28	“A few days after 1 dose”	SPTCL	Ad26 viral-vector-based	Injection site, upper arm	Cyclosporine plus prednisone
7	F/80	1 day after 1 dose	EMZL	BNT162b2	Right temporal mass	No treatment
8	M/76	10 days after the booster dose	PC-ALCL	mRNA-1273*	Right arm upper-external surface 6 cm	No treatment

Table reproduced from Cavanna et al., *Medicina*, 2023. © MDPI. Abbreviations: ALCL: anaplastic large-cell lymphoma; DLBCL: diffuse large B-cell lymphoma; EBV: Epstein-Barr virus; EMZL: extranodal marginal zone lymphoma; PC-ALCL: primary cutaneous anaplastic large-cell lymphoma; SPTCL: subcutaneous panniculitis-like T-cell lymphoma. *The two previous vaccination doses were BNT162b2.)

one day after receiving a second dose of the BNT162b2 mRNA COVID-19 vaccine (Pfizer–BioNTech, USA). Ophthalmologic exam revealed secondary angle-closure glaucoma, bullous retinal detachment, and extensive intraocular hemorrhage. Ocular imaging and confirmed magnetic resonance imaging (MRI) revealed an ill-defined heterogeneous subretinal lesion, with histopathology confirming necrotic uveal melanoma (Figure 3G). Gullotti et al. [55] also described an otherwise healthy 53-year-old man who presented with ipsilateral axillary lymphadenopathy and associated discomfort shortly after receiving a COVID-19 vaccine. Fine-needle aspiration performed within two months of vaccination revealed metastatic melanoma, and subsequent 18F-FDG PET/CT imaging demonstrated intensely hypermetabolic axillary and supraclavicular lymphadenopathy without identification of a primary tumor (Figure 3H).

Glioblastoma

Tosun et al. [29] reported a 40-year-old man presenting with left hemiparesis. He had received COVID-19 vaccination 3 weeks before. Brain MRI showed a central cystic necrotic lesion with indistinct borders in the right frontal lobe as mild peripheral contrast enhancement surrounded by smaller nodular lesions.

O Sullivan et al. [84] also describe a 31-year-old female who first noted a slight weakness of her right leg about 7 days after receiving the first dose of a COVID-19 mRNA vaccine (Comirnaty®BioNTech Manufacturing GmbH, Germany). She initially reported slight drowsiness and headache without fever following vaccination, which resolved within 24 h. Following the administration of the second intramuscular dose of the vaccination, 21 days after the first, the preexisting weakness of the right leg rapidly worsened and was accompanied by severe headache and night chills. Neurological examination on day 28 showed a mild central paresis of the right leg and numbness of the plantar surface of the foot (Figure 3I).

Other

Kim et al. [31] describe two cases of gastrointestinal polyposis (Cronkhite–Canada syndrome) shortly after administration of an mRNA booster vaccine for COVID19. Both showed numerous erythematous gastric and colonic polyps with villous atrophy throughout the small intestine (Figure 3J). The authors note that the timing, autoimmune features, and steroid responsiveness raise the possibility that mRNA vaccination may trigger Cronkhite–Canada syndrome in genetically susceptible individuals, warranting clinical vigilance. Sasa et al. [33]

report on axillary lymphangioma following COVID-19 in a Japanese woman in her 80s who received a second injection of the Pfizer-BioNTech COVID-19 vaccine in her left deltoid muscle in 2021 (Figure 3K). She had a history of right breast cancer (T1N0M0) and had undergone breast-conserving surgery and sentinel node biopsy in her 70's. Postoperative follow-up examinations were continued, and no sign of recurrence, including in the left axial region, was observed until 2021. There was no evidence of trauma to the left axial region. Her early adverse reaction following vaccination was mild pain at the inoculation site on the day of vaccination and the following day. However, 3 months after the second vaccination, she noticed left axillary swelling.

***De novo* disease versus recurrence or progression**

Most publications, including all the examples above, described *de novo* malignancies or apparent “unmasking” or activation of previously subclinical disease. A smaller subset focused predominantly on recurrence, progression, or metastatic reactivation in patients with a documented cancer history. An additional 13 publications reported mixed cohorts, including both new diagnoses and recurrences or provided explicit quantification of both categories. Only one publication did not clearly distinguish between new-onset and recurrent disease.

Taken together, these patterns indicate that the observed signal in the literature is not restricted to recurrence or flare of known malignancies. Rather, a substantial proportion of reports involve first-time cancer diagnoses temporally associated with COVID-19 vaccination or SARS-CoV-2 infection, highlighting the need to evaluate potential mechanisms that could contribute to disease initiation, unmasking, or acceleration.

Timing of onset

Across the included studies, the timing of cancer onset following COVID-19 vaccination varied substantially, indicating that latency was not confined to a single early window. Approximately half of the case reports described diagnoses occurring within 2–4 weeks of vaccination, with some reported as early as 7–14 days. However, many reports also documented longer intervals, including diagnoses at 2–3 months, 4–6 months, and beyond eight months after vaccination. Importantly, reports with short intervals are inherently more likely to be recognized and published as temporally notable.

In addition, in many reports describing diagnoses within the first month, the event occurred after a second dose or booster, complicating attribution to any specific exposure and precluding definition of a uniform latency period. Multicenter analyses frequently characterized

latency as variable, spanning weeks to months, and several reviews or population-level studies reported mean onset intervals of approximately 8–9 weeks.

Tumor growth rates vary significantly among tumor types from the fastest growing lymphomas and leukemias to slower growing solid tumors [87–92]. Accordingly, while a subset of published cases report diagnoses within weeks of vaccination, the broader literature reflects a continuum of reported latencies over several months, often in the context of cumulative exposure. These observations are therefore best interpreted as descriptive and hypothesis-generating, underscoring the need for standardized latency definitions and systematic evaluation in appropriately controlled studies.

Population-level and registry-based studies

Three large-scale population-level analyses provided broader epidemiologic context to complement the case-based literature. Two retrospective population-level investigations, one in Italy [20] and one in South Korea [8], quantified cancer incidence and mortality trends among vaccinated populations. Kim et al. [8] analyzed approximately 8.4 million individuals between 2021 and 2023 to assess 1-year cumulative cancer incidence following COVID-19 vaccination using the South Korean National Health Insurance Service database. The authors reported statistically significant associations between vaccination and six specific cancers, including thyroid (HR 1.35), gastric (HR 1.34), colorectal (HR 1.28), lung (HR 1.53), breast (HR 1.20), and prostate cancer (HR 1.69) using propensity score matching and multivariable Cox proportional hazards models. Associations varied by vaccine platform, with mRNA vaccines linked to thyroid, colorectal, lung, and breast cancers, and cDNA/adenoviral vaccines associated with thyroid, gastric, colorectal, lung, and prostate cancers; heterologous vaccination was associated with thyroid and breast cancer. Stratified analyses suggested effect modification by sex and age, and booster-dose analyses identified increased risks for gastric and pancreatic cancer. The authors emphasized that despite adjustment for measured confounders, residual confounding, detection bias, and limited follow-up preclude causal inference, and that the findings should be interpreted as epidemiologic associations warranting further study rather than evidence of vaccine-induced cancer risk.

Acuti Martellucci et al. [20] evaluated associations between SARS-CoV-2 vaccination, all-cause mortality, and hospitalization for cancer using multivariable Cox proportional hazards models in a population-wide retrospective cohort study of 296,015 residents of the Pescara province in Italy followed for up to 30 months (June 2021–December 2023). Hospitalization for cancer of any site was found to be modestly higher among vaccinated individuals compared to unvaccinated (≥ 1

dose: HR 1.23, 95% CI 1.11–1.37; ≥ 3 doses: HR 1.09, 95% CI 1.02–1.16), with site-specific increases observed primarily for colorectal (HR 1.35), breast (HR 1.54), and bladder cancer (HR 1.62) after ≥ 1 dose, and for breast and bladder cancer after ≥ 3 doses. These associations varied by sex, prior SARS-CoV-2 infection, vaccine type, and lag time between vaccination and outcome, and were attenuated or reversed when a longer minimum latency of 365 days was applied. The analyses adjusted for age, sex, prior SARS-CoV-2 infection, and multiple comorbidities (including cardiovascular disease, diabetes, COPD, kidney disease, and prior cancer), but lacked information on key behavioral and healthcare-utilization confounders such as smoking, cancer screening, and healthcare-seeking behavior. The authors explicitly note that residual confounding, healthy-vaccine bias, detection bias, and reliance on hospitalization data as a proxy for cancer incidence limit causal interpretation, and they characterize the findings as preliminary and hypothesis-generating rather than evidence of vaccine-induced cancer risk. Both studies provide early, population-level associations rather than causal estimates and underscore the importance of long-term follow-up and molecular correlation to distinguish true biological effects from health-system or behavioral confounders.

In addition to these population-level studies, a recent US Armed Forces Health Surveillance Division (AFHSD) report was also identified that presented population-level analyses of non-Hodgkin lymphoma (NHL) incidence among active-duty U.S. service members from 2017 through 2023 [85]. The U.S. Department of Defense (DoD) mandated COVID-19 vaccination for all active-duty service members (~1.3 million) beginning in late 2020, with near-universal compliance achieved by

mid-2020; this cohort offers a rare longitudinal view of cancer incidence across this transition. Using data from the Defense Medical Surveillance System (DMSS), the authors calculated annual incidence rates (IRs) per 100,000 person-years and categorized cases by lymphoma subtype and the 2017–2020 interval largely represents a pre-vaccine baseline, whereas 2021–2023 reflects a fully vaccinated, post-pandemic cancer incidence [86] (Figure 4).

Notably, a rise in mature T/NK-cell lymphomas began across the 2020–2021 transition which spans the period of COVID-19 infection and the beginning of widespread vaccination in the military. Beginning in 2021, a ~50% increase in specified/unspecified and non-follicular NHL subtypes, accompanied by a persistently elevated incidence of mature T/NK-cell lymphomas relative to pre-pandemic years was observed. Notably, the authors did not attribute the observed changes in NHL incidence to vaccination or infection, and the analysis was not designed to establish causality at the individual level. Changes in diagnostic practices, healthcare access and utilization, and pandemic-related disruptions to routine medical care cannot be excluded from this time-trend analysis as with others conducted during the pandemic period. However, these findings provide descriptive temporal trends within a unique and highly structured population, providing an epidemiologic framework for future controlled analyses.

Taken together, these population-level analyses combined with the case-based literature indicate that a cancer signal warrants further prospective evaluation to determine whether COVID-19 vaccination confers any measurable cancer risk or merely reflects surveillance and reporting biases.

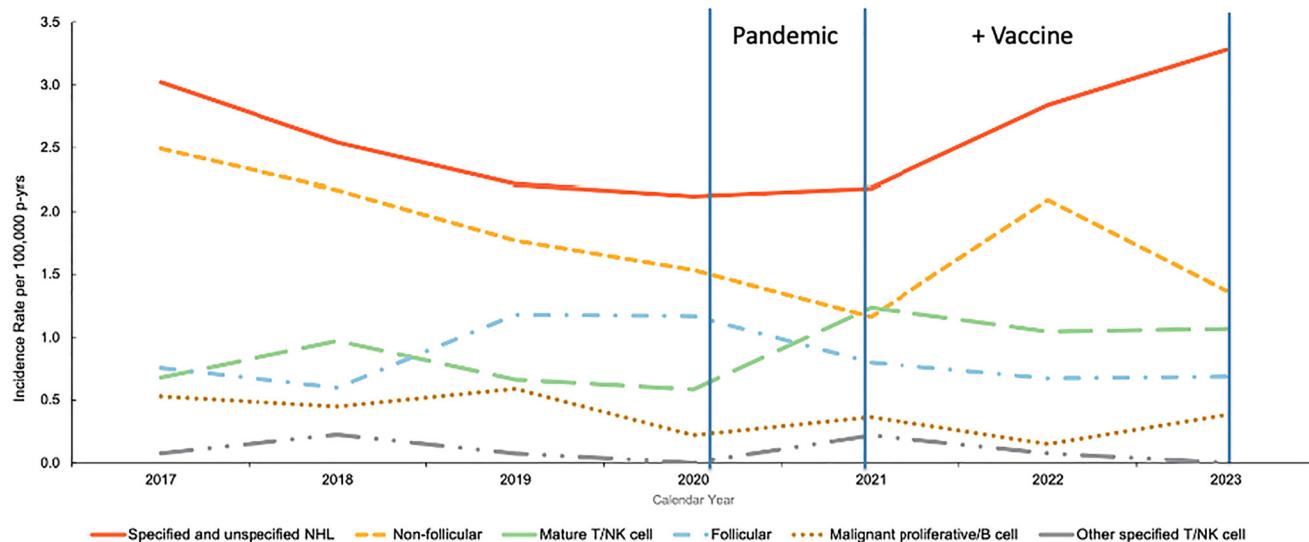


Figure 4: Annual incidence rates of non-Hodgkin lymphoma (NHL) subtypes among active-component U.S. service members, 2017–2023. Figure adapted from Russell et al. [85] using Defense Medical Surveillance System data demonstrating rise in specified/unspecified NHL and mature T/NK-cell subtypes. Vertical lines denote key timepoints: the onset of the COVID-19 pandemic (early 2020) and the beginning of the Department of Defense vaccine mandate (late 2020–early 2021).

MATERIALS AND METHODS

A comprehensive search of the world's literature was conducted using PubMed, Scopus, Web of Science, Google Scholar, and React19 between January 2020 and April 2025. Eligible publications included case reports, case series, cohort or population-level analyses, systematic reviews, and mechanistic or preclinical studies that described either (i) new-onset, recurrent, or rapidly progressive malignancy temporally associated with COVID-19 vaccination or SARS-CoV-2 infection, or (ii) experimental evidence implicating vaccine or infection-induced immune perturbations in oncogenic, proliferative, or metastatic processes.

Initial searches in PubMed using conventional keyword combinations such as "COVID-19 vaccine and cancer," "vaccination and cancer," "COVID-19 vaccine and tumor," or cancer-specific terms paired with "COVID-19 vaccine" yielded little to no indexed results. Even when known case reports were used as anchors for "similar articles," PubMed returned no related entries. This highlighted a structural limitation in standard indexing pathways and necessitated a broader, more strategic search approach.

A general web-based search (e.g., Google) returned an autogenerated AI summary when queried for the terms "COVID vaccine and cancer" indicating that major health agencies, including the Centers for Disease Control and Prevention (CDC) and the National Cancer Institute (NCI), recommend COVID-19 vaccination for individuals with cancer and assert that the vaccines are considered safe for this population and are not believed to cause cancer or precipitate recurrence. Therefore, an expanded search strategy was implemented using combinations of general and tumor-specific terms, including: "COVID-19," "SARS-CoV-2," "spike," "vaccination," "vaccine," "tumor," "cancer," "neoplasia," "malignancy," "recurrence," "progression," "lymphoma," "leukemia," "melanoma," "glioma," "adenocarcinoma," "sarcoma," "Kaposi," "Merkel cell," "cardiac", and related descriptors. Databases were searched using Boolean operators, varied term order, and MeSH/non-MeSH variants to overcome incomplete tagging or atypical indexing of case reports.

Studies were included irrespective of patient age, sex, geographic region, cancer histology, or vaccine platform (mRNA, viral-vector, or inactivated). Exclusion criteria consisted of commentaries, opinion correspondence, purely theoretical articles lacking clinical or experimental data, and duplicate case entries across publications. Studies labeled as "COVID-associated" or "COVID-related", particularly for cardiac tumors ultimately described patients who tested negative for SARS-CoV-2 [93]. For methodological consistency, we excluded such reports from the infection-focused section of the analysis, as the absence of virologic confirmation

precludes attributing the observed malignancy to active or recent infection. Reference lists of systematic reviews and larger case compilations were manually screened to identify secondary citations not captured in the primary search. All included articles were independently cross-referenced in PubMed when possible, to confirm indexing status and ensure completeness.

Mechanistic hypotheses linking COVID-19 vaccination or infection to oncogenic events

The case studies and emerging population-level data described above may represent an early signal of a possible association between vaccination or infection and cancer that warrants further investigation. This raises the question: if there is an association, what might be the mechanistic basis for it?

Viruses can cause cancer [94–97]. The relationship between viral infection and cancer has been well-documented for Human Papilloma Virus (HPV) that causes cervical cancer, head and neck cancer, as well as anal cancer that is increased among HIV-infected individuals. Hepatitis B Virus (HBV) and Hepatitis C Virus (HCV) cause liver cancer. Epstein Barr Virus (EBV) causes nasopharyngeal cancer, Burkitt's Lymphoma, and other cancers. The human herpes virus KSHV/HHV-8 causes Kaposi's sarcoma, the Human T-cell Leukemia Virus (HTLV-1) causes adult T-cell leukemia or lymphoma, and the Merkle Cell Virus (MCV) causes Merkle cell skin cancer. Several viruses are suspected of causing cancer including SV40 (mesothelioma, primary brain and bone cancers, among others) and HCMV (glioblastoma, medulloblastoma, breast, colon and prostate cancer). HIV is strongly associated with Kaposi's sarcoma, cervical cancer, lymphoma, anal cancer, and other malignancies, largely through immunosuppression and co-infection with oncogenic viruses. It has been known for decades that viral proteins target host tumor suppressors such as p53 and Rb, suppress the immune system, and activate oncogenic signals.

In addition, the COVID mRNA vaccines work by instructing the target cells to produce the SARS-CoV-2 spike protein. This occurs by introducing a synthetic, modified mRNA (mod-mRNA) which incorporates non-natural pseudouridine into its coding region to prolongs the stability of the mRNA beyond that of natural mRNA. Introduction of the mod-mRNA is accomplished using lipid-based transfection in the form of lipid nanoparticles (LNPs). The result is highly efficient transfection of the mod-mRNA into target cells with biochemical and pharmacological behavior different from naturally occurring mRNA. Consequently, the mod-mRNA is transcribed into the foreign spike protein (as well as other frameshifted protein products), which elicits a robust immune response [98–102]. Given the stability of pseudouridine modified mRNA, along with the residual

DNA in the mRNA vaccine formulations [103–108], the mRNA vaccines are delivering exogenous genetic material (DNA and RNA (in the form of engineered nucleic acids)) into a patient's cells. The COVID19 mRNA vaccines produce Spike protein that is encoded by a stable mRNA and has been found to be long-lived in the human body [109, 110]. These nucleic acid elements have been reported to contribute to Post-Covid Vaccine Syndrome (PCVS/ PVS) [110, 111]. Thus, these vaccines fit the definition of gene therapy [112, 113]. Despite this, there are efforts by the EU to modify the definition of gene therapy to exempt mRNA vaccines from this category [114].

While there are no studies demonstrating a direct causal mechanism by which the mRNA vaccines induce cancer, cumulative molecular effects from persistent spike protein [115, 116], the immune activation and inflammation from repeated vaccination [117–119], or the potential for genomic integration events [120] might contribute to events that could in theory manifest in cancers following vaccination or infection. Given the rapid onset of aggressive and rare tumors from the literature, cancers arising weeks to months after vaccination would be perhaps more consistent with mechanisms involving tumor promotion rather tumor initiation *per se*. However, mechanisms involving initiation are also considered. Here we present least three biologically plausible mechanisms that might explain an association between COVID-19 vaccination and cancer; two of them overlapping with covid infection, immune dysfunction and spike protein biology, and reactions due to DNA impurities restricted to vaccination.

Immune dysregulation

The rapid appearance of cancer, the anatomical proximity of the tumors to vaccine sites, and the histologic signatures of inflammation the support immune mechanisms that promote the progression of latent clones rather than *de novo* carcinogenesis. We hypothesize two interrelated processes: localized inflammation and modulation of the tumor microenvironment with transient functional immunosuppression that relaxes immune surveillance. Might account for hyperprogression of latent or occult cancer cells (Figure 5).

Numerous studies (both human and animal) have shown that COVID mRNA vaccines and infection trigger production of proinflammatory cytokines including interleukin-6 (IL-6), TNF- α , and IL-1 β within 1–3 days after vaccination [121–124]. In the case of vaccination, the reaction is due to the innate immune response to the mRNA and lipid nanoparticle (LNP) components, which activate pattern-recognition receptors TLR7/8 and NLRP3 [125–128]. Therefore, local production of these cytokines will occur wherever the mRNA and LNPs are biodistributed, which include the injection site, draining lymph nodes, as well as other distant sites [129].

IL6 activates STAT3 which drives cancer cell proliferation, survival, angiogenesis, and immune suppression in the tumor microenvironment [130]. TNF- α activates NF- κ B and AP-1 that also drives cell survival, proliferation, angiogenesis, and immune evasion. TNF- α can create a self-sustaining inflammatory loop by recruiting myeloid-derived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), and regulatory T cells (Tregs) that suppress cytotoxic T-cell activity and produce additional TNF- α , IL-6, and IL-10. TNF- α induces further expression of IL-6, CXCL1/2/8, and COX-2, which fosters further proliferation, angiogenesis, and immune evasion [130]. IL-1 β upregulates VEGF, MMPs, and integrins, promoting neovascularization and extracellular matrix remodeling. IL-1 β drives polarization of macrophages toward the M2-like (tumor-promoting) phenotype, expands Th17 cells and an neutrophils, thereby contributing to chronic inflammation and immunosuppressive tumor microenvironment [130].

Together IL-6, TNF- α , and IL-1 β constitute a synergistic pro-inflammatory circuit capable of stimulating proliferation and angiogenesis. Together, this circuit is known to rapidly promote the development of cancer if transformed or pre-malignant cells already exist. Indeed, there are several reports demonstrating acceleration of pre-existing disease or reawakening from dormancy following inflammation [74, 131–134]. A synchronized surge of these three cytokines could provide a coordinated inflammatory storm that converts indolent or dormant transformed cells into rapidly proliferating, and angiogenic, malignancies (Figure 5). Therefore, research is needed to better understand whether the hypothesis that an inflammatory cytokine cascade is unleashed by mRNA vaccination could contribute to or even lead to post-vaccination cancer events.

In addition to influencing the behavior of pre-existing neoplastic cells, this transient cytokine-driven inflammatory surge could also modulate antiviral or antitumor immune surveillance. IL-6, TNF- α , and IL-1 β recruit immunosuppressive myeloid populations and expand regulatory T cells, while dampening cytotoxic T-cell activity, functions that are essential for maintaining control of latent oncogenic viruses such as EBV, HHV-8, and MCV [135–137]. Accordingly, the observation that several post-vaccination cases involved virus-associated cancers (EBV, HHV-8, MCV) raises the possibility that short-lived alterations in immune surveillance may permit episodic viral reactivation or progression of virus-driven tumors. Mechanistically, transient impairment of cytotoxic T-cell surveillance prevents reactivation and replication of latent oncogenic viruses, leading to expression of viral oncogenes and proliferation of infected host cells. Similar processes are well documented in states of clinical immunosuppression, or hyperprogression in patients receiving immune checkpoint inhibitors [138–140].

Beyond innate cytokine induction, several studies have described transient adaptive immune changes following mRNA vaccination or acute infection [110, 141–145]. Although these findings are generally interpreted as reflecting physiological immunoregulation rather than overt immune dysfunction, they may correspond to short periods of reduced immune responsiveness [146–149]. If such windows occur, whether systemically or localized to specific tissue niches, they could, in theory, allow transient expansion of latent viral or neoplastic clones. At present, whether transient functional immunosuppression occurs after mRNA vaccination, and whether it is localized or systemic, remains unclear. Direct studies are needed to determine whether innate cytokine induction and adaptive immune modulation could contribute to or influence post-vaccination cancer events in susceptible individuals.

Evidence supporting these mechanisms have been suggested after SARS-CoV-2 infection itself. For example, Gregory et al. [74] reported aggressive glioblastoma at a median of 35 days following COVID-19 infection, an in one patient documented COVID-19 vaccine prior to diagnosis. The rapid onset of glioblastoma was attributed to immune disruption which may be in part due to the neurotropism of SARS-CoV-2 [150, 151] or the biodistribution of the LNPs [125, 127, 129] and the immune response to this. Indeed, Spike protein as has been localized to brain tissue as well as glioblastoma cells and macrophages surrounding the tumor cells [87]. Hu et al. [72] similarly demonstrated that direct viral exposure, and the presence of Spike protein, with cytokine-mediated injury in glioma organoids enhanced tumor cell proliferation and invasiveness, supporting a model of infection-driven tumor stimulation. For breast cancer, Chia

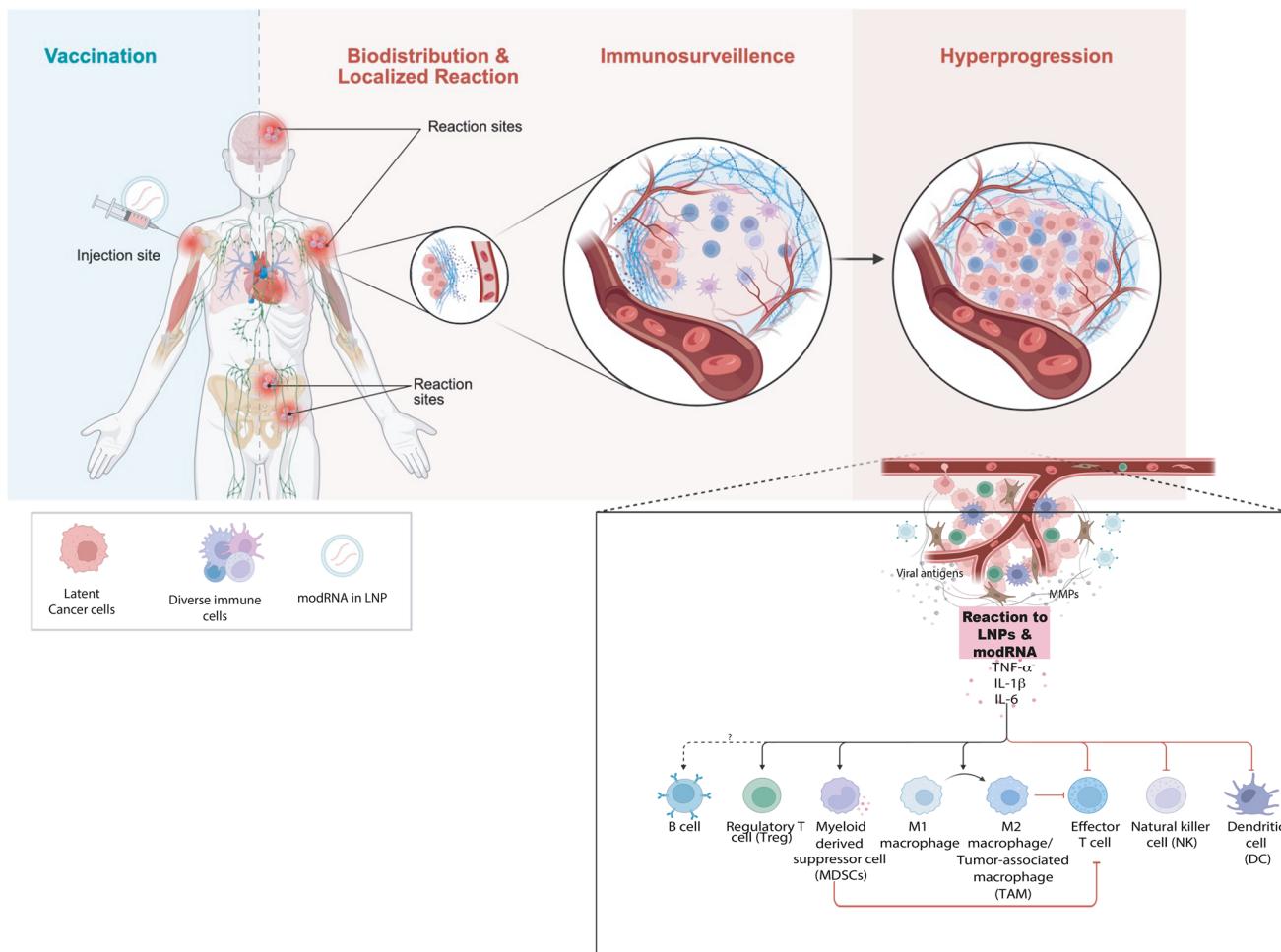


Figure 5: Proposed mechanism of tumor hyperprogression following COVID-19 vaccination. (A) Conceptual model illustrating how inoculation with mRNA vaccine leads to immune reactions depending on its biodistribution. Strong immunostimulation can override immunosurveillance of latent cancer cells and trigger tumor hyperprogression. (B) Schematic representation of the major immune cell types influencing tumor growth and immune regulation following mRNA vaccine exposure. LNP-encapsulated modified mRNA (modRNA/mRNA) interacts with innate immune sensors altering cytokine signaling (TNF- α , IL-1 β , IL-6) and immune-cell polarization leading to immunosuppression and reduced cytotoxic CD8 $^{+}$ T-cell activity. Expansion of myeloid suppressor populations, along with pro-tumor cytokine feedback loops, fosters accelerated tumor cell proliferation and immune evasion. The imbalance between anti-tumor (M1, CD8 $^{+}$, NK) and pro-tumor (M2, Treg, MDSC) networks favors tumor hyperprogression.

et al. [74] showed that respiratory viral infections including SARS-CoV-2, could rapidly awaken dormant metastatic breast cancer cells in the lung through interferon-driven activation of the STAT1–NF-κB axis, which remodels the local niche into a pro-metastatic state. Both the virus and the mRNA vaccines engage innate immune sensors and elicit complex cytokine and interferon responses that can remodel the tumor–immune interface.

Together, these observations suggest that acute inflammatory activation, short-lived immune refractoriness, and transient lapses in cytotoxic surveillance form a biologically plausible framework through which vaccination could influence and promote the behavior of pre-existing or latent neoplastic cells. The immunological mechanism suggests that infection and vaccination may operate along a common biological continuum, differing primarily in intensity, biodistribution, and persistence of the immune and molecular perturbations they induce. Although direct causal evidence is not yet available, the convergence of innate cytokine surges, transient modulation of T- and B-cell dynamics, and signals associated with immune regulation highlights the importance of further study. Clarifying the magnitude, duration, and tissue specificity of these post-vaccination immune states will be essential for determining whether, and in which individuals, they have clinical relevance for cancer progression or recurrence.

Spike protein biology

The transformation of a normal cell into a cancer cell involves disruption of multiple safeguards controlling cell growth, survival, and DNA repair. Laboratory studies have reported that the spike protein, whether it is produced by infection or by vaccination, has biological activities [110, 145, 152–158] with oncogenic potential [159–161]. For example, in addition to interacting with ACE2 receptors, spike protein fragments have been shown to interact with NRP-1, integrins, and TLRs leading to VEGF/NRP-1 signaling [155, 162, 163]. Spike protein has also been reported to induce DNA damage [160, 164, 165] and modify p53 pathway under oxidative stress [164, 166]. Therefore, in theory, such interactions of spike protein with these pathways could contribute to cellular transformation, both from the vaccine but also from infection, especially if the spike protein remains present long after vaccination or from multiple COVID infections.

The spike protein produced by the vaccines has been reported to persist for weeks, months and even years [109, 110, 116, 167–170] after vaccination, providing potential long-term activity in cells. Moreover, the stabilized Spike protein produced by the covid vaccines (Spike-2P), differs from the natural protein in SARS-CoV-2 as it contains two proline substitutions (K986P and V987P) that enable stabilization [171]. Because of this, it will be important to assess whether cancer incidence correlates with the variant spike protein expression (or persistence) in the body but

also whether it is this version that is present in tumors as has been reported in glioma and astrocytoma [73] as well as metastatic breast cancer [36]. In a recent case report of metastatic breast cancer to the skin, the lesion appeared within one month after the 6th dose of vaccination (Pfizer-BioNTech) and the metastatic cancer cells stained for spike protein, but not for nucleocapsid protein of SARS-CoV-2 virus ([36], Figure 3F) suggesting it was vaccine-derived spike protein. Hence, chronic exposure to an agent with biological activity that disrupts cell cycle and DNA damage response pathways could represent a novel etiological factor to cancer. Of note and relevance to glioblastoma or other central nervous system (CNS) pathologies after either COVID infection or vaccination could be the CNS-tropism of spike protein [150, 151, 172].

DNA contaminants

Residual DNA in biologics is a well-established and acknowledged byproduct of vaccine manufacturing, with limits set forth by the FDA and World Health Organization (WHO), but only for naked DNA, not LNP encapsulated DNA [173]. The DNA impurities in mRNA vaccines arise due to the byproduct of in-vitro transcription [174], and can include double strand DNA (dsDNA), abortive RNAs and RNA:DNA hybrids [103, 174]. They are encapsulated by nanolipids allowing for more stable and efficient access into cells increasing the risk of integration [128, 175, 176]. Furthermore, the residual DNA in the mRNA vaccine formulations [103–108] from the manufacturing process exceed the established limits even for naked DNA. Studies have directly compared the transfection efficiency of naked DNA to LNP encapsulated DNA and shown that integration of lipid-based transfection is significantly higher than naked DNA [175]. Moreover, skeletal and cardiac muscles are well known to take up (and even express naked plasmid DNA) *in vivo* [177–179]. Notably, a study of cardiac tumors in the post-COVID period revealed both a 1.5% increase in tumor incidence and the expression of spike protein with the tumors, particularly in those associated with vaccination [86].

The quantity of residual DNA reported in several independent assessments exceeds recognized limits for naked DNA, and the size distribution of DNA fragments, when combined with enhanced transfection efficiency due to LNPs raises the possibility of genomic insertion. In addition, because SV40 regulatory elements are present in the BNT2b vaccine impurities [180], when inserted into genome this DNA can alter the expression of adjacent sequences and/or normal gene regulation and increases tumorigenic potential [120, 181]. Foreign DNA, especially when delivered in the highly inflammatory LNPs [182] can activate innate immune sensing pathways, including the cytosolic cGAS–STING and endosomal Toll-like receptor 9 (TLR9), leading to type I interferon and inflammatory cytokine responses [183, 184].

The limits on DNA impurities were established for naked DNA [185], not LNP-encapsulated DNA which causes enhanced cellular uptake and intracellular persistence of DNA fragments. This will increase the opportunity for insertional mutagenesis leading to possible genomic rearrangements, as well as integration and expression of persistent spike protein, disruption of normal gene regulation, as well as possible activation of proto-oncogenic pathways, or inactivation of tumor suppressors. In fact, *in vitro* studies demonstrate genomic integration rates of ~1–10% of initially transfected cells with lipid-based delivery systems [120]. No studies have been conducted demonstrating that the level of DNA impurities present in the vaccines are insufficient to transfect cells, nor have studies ruled out the possibility of integration.

Gaps in knowledge

Despite the unprecedented global scale of COVID-19 vaccination, profound gaps remain in our understanding of how mRNA vaccine platforms interact with fundamental pathways of host defense, tissue homeostasis, and tumor biology. These gaps span molecular, cellular, organismal, and population level biology. There is an absence of data linking mRNA vaccines, and especially COVID-19 mRNA vaccines to downstream biological consequences.

At the molecular level, major knowledge gaps concerning how chemical and structural modifications of the SARS-CoV-2 spike protein, nucleoside and amino acid substitutions (e.g., N¹-methylpseudouridine), and LNP formulations influence host-cell signaling, genomic stability, and immune regulation. Vaccine engineering has focused on maximizing antiviral immunogenicity, yet far less is known about potential collateral interactions between spike protein expression, tumor-suppressive, DNA damage or stress response pathways that could be inadvertently modulated during intense immune activation or altered cellular signaling that impacts on host defenses against cancer.

The distinction between vaccination associated tumor initiation and promotion also remains unresolved. There is no empirical validation that vaccination only accelerates pre-existing disease rather than also initiating new malignancies. Because somatic mutations and dormant neoplastic cells are ubiquitous in adult tissues, short-latency tumor emergence over weeks to months may reflect the promotion of latent clones rather than de novo carcinogenesis, a phenomenon consistent with hyperprogression observed in subsets of patients receiving immune checkpoint inhibitors [138, 139]. Oncogenic drivers such as MDM2/MDM4 amplification or EGFR amplification, overexpression, or mutations have been implicated in hyperprogression [138, 140, 186] and metastatic aggressiveness [187, 188], suggesting that

vaccine-induced cytokine or checkpoint shifts could theoretically converge on similar oncogenic pathways. Also unclear is how mRNA vaccines targeting SARS-CoV-2 might be sensitizing tumors to immune checkpoint inhibitors as has been recently suggested [19].

Cellular and immunologic gaps

At the cellular level, there is limited mechanistic understanding of how mRNA vaccine components, spike protein persistence, or repeated immune activation shape innate–adaptive immune crosstalk, particularly in dendritic cells, macrophages, and stromal compartments. The molecular triggers and long-term immune consequences of a hyperinflammatory response observed in both severe COVID-19 and rare post-vaccination events [189] remain poorly characterized. Equally unexplored is the role of the microbiome in modulating vaccine responsiveness, systemic inflammation, and tumor promotion. Understanding how antigen persistence, cytokine polarization, and pattern-recognition receptor activation influence local tissue remodeling, cellular senescence, and pro-tumorigenic inflammation represents a critical unmet need.

Host susceptibility and biodistribution

At the organismal level, the greatest uncertainty lies in the heterogeneity of host susceptibility. For example, the heterogeneity of individual differences in baseline state of activation, responsiveness, and regulation in response to COVID-19 vaccination has been reported [190]. Furthermore, heterogeneity of DNA repair, epigenetic plasticity, and cytokine response level is also not well understood and likely modulates vaccine response and risk. Variable LNP biodistribution, including uptake in liver, spleen, bone marrow, and lymphoid tissues, may alter both immune potency and potential off-target effects, yet these parameters have not been systematically profiled in humans. There is also a lack of data on how vaccination during or shortly after SARS-CoV-2 infection, or cumulative exposure to multiple mRNA doses, affects long-term immune homeostasis and tumor surveillance. Interactions between vaccine-induced inflammation and latent oncogenic viruses (e.g., EBV, HHV-8, MCPyV) remain particularly underexplored.

Population and epidemiologic gaps

At the population level, large-scale epidemiologic studies remain limited and often inconclusive. Existing registries rarely integrate molecular or immunologic correlates, hindering causal inference. Moreover, current pharmacovigilance systems were not designed to detect rare but biologically informative oncologic events,

creating a blind spot between individual case reporting and aggregated safety analyses. Robust longitudinal follow-up of vaccinated cohorts with integrated molecular profiling will be essential to distinguish true biological signals from background incidence and to identify susceptible subgroups.

Alternative preventive and therapeutic strategies

Addressing key knowledge gaps will require exploration of complementary antiviral strategies that reduce infection risk while minimizing host perturbation. For example, MEK inhibitors have been shown to suppress ACE2 expression and viral entry [191], yet host-directed antivirals have received little attention. Likewise, enhancing innate immunity through agents that boost pattern-recognition receptor signaling or interferon responses could theoretically mitigate both COVID-19 infection and cancer risk by restoring balanced immune surveillance. Indeed, there are approaches validated by FDA of approved immunomodulators [192] and emerging analogs [193].

Major gaps exist in forensic evidence to evaluate the spread of COVID infection or COVID vaccine Spike protein into cancer tissues as well as effects on cellular growth control pathways *in vivo*. We currently do not have tissue-level, mechanistic, or molecular studies that trace where spike protein localizes in the body after infection or vaccination, especially in tumors. No systematic research has mapped spike protein distribution within tumors (solid or hematologic), either through immunohistochemistry, mass spectrometry, RNA *in situ* hybridization, or other molecular tissue-tracking methods. There have been essentially no *in-vivo* studies (animal or human tumor samples) showing how spike protein exposure affects proliferation, signaling, apoptosis pathways, tumor-immune interactions, or oncogene/tumor suppressor regulation.

In sum, the field faces interlocking knowledge gaps encompassing incomplete molecular characterization of vaccine–host interactions, insufficient mechanistic study of immune dysregulation and tissue remodeling, poorly defined host susceptibility factors, and limited longitudinal surveillance. Bridging these gaps will require multi-scale, interdisciplinary research.

Limitations

The findings of this review should be interpreted within several important limitations inherent to the available literature and study design. First, although there are larger studies published, most reports are single-patient case descriptions or small case series. Many reports lack documentation of pre-existing conditions, prior oncologic history, concurrent infections, or medications that could confound interpretation. Therefore, while these

observations are valuable for early signal detection, they are highly susceptible to publication bias and selective reporting. It is plausible that cases perceived as unusual or temporally linked to vaccination are preferentially submitted for publication, while the absence of comparable control observations limits inference regarding incidence or relative risk.

Second, the heterogeneity of the studies that span both mRNA and non-mRNA vaccine platforms, doses, populations, and diagnostic standardization introduces important variabilities when it comes to the associations between mRNA vaccines and other COVID-19 vaccines. The mechanistic hypotheses proposed here remain speculative in the absence of direct *in vivo* validation. No current studies have demonstrated oncogenic transformation or tumor initiation causally attributable to the COVID mRNA vaccine or its components. Nor have animal studies demonstrated vaccine-induced tumor promotion either. These mechanisms should therefore be regarded as biologically plausible models that warrant targeted experimental study.

Finally, this literature review relied on publicly available literature that lacked standard medical subject headings (MeSH) indexing for peer-reviewed papers. The results of this review demonstrate that there indeed exists a body of literature on this subject matter across many journals that not well indexed or cross-referenced using obvious indexing vocabularies that prevent them from being retrieved by conventional database queries. This imposes inherent limitations on both data completeness and verification but also in identifying knowledge gaps.

Ethical considerations

Different disciplines adopt different standards for evidence used to determine standard of care that should be shared widely. In 2025, there is a major divide about published evidence that doesn't reach a certain standard at a population level being viewed by some as "misinformation," or "fearmongering." In the field of Oncology, there is a standard that has been in place for decades regarding how adverse events are viewed in the process of drug development as well as in clinical practice. A single serious adverse event is reportable to institutional review boards (IRB's) and regulatory agencies such as the FDA. The mRNA vaccines incorporate mechanisms commonly associated with gene-therapy technologies, though their regulatory review followed the vaccine pathway rather than the criteria normally applied to human gene-therapy products, which rigorously evaluate. Cancer (including insertional mutagenesis, clonal expansion, leukemogenesis, and treatment-related malignancy) is a key safety endpoint that gene-therapy regulations are explicitly designed to monitor, but vaccines are not.

Regarding COVID mRNA vaccines, in the US and globally, there is no informed consent that captures all known and potential adverse events. All known and potentially serious adverse events are required to be disclosed in vaccine information sheets, informed consent forms, and it is important for physicians and other health care providers to be well-educated concerning risks regardless of how rare they are. An important example that led to practice change in the US after decades which lacked DPYD testing. This now mandated test identifies pathogenic or reduced-function variants in the DPYD (dihydropyrimidine dehydrogenase) gene that impairs 5-FU/capecitabine metabolism. Changes in practice occurred recently and has become in line with European practice to reduce the risk of severe toxicity from exposure to 5-fluorouracil chemotherapy.

Cancer susceptibility varies among individuals in the population as governed by genetic and environmental factors and a growing body of literature adds socioeconomic factors that interact with the other two factors in complex ways. It is likely that the susceptibility to cancer following COVID vaccines varies greatly within the population with some individuals at greater risk. It is important to recognize these issues and to study them in order to develop improved guidance for a risk: benefit analysis in the setting of informed consent.

The long timeline required to establish or exclude causality in oncology is often measured in years or decades. This should not lessen the immediate ethical responsibility to provide accurate and current information to individuals considering vaccination or additional boosters. The fact that definitive causal inference demands mechanistic studies, longitudinal cohorts, and large epidemiologic analyses should not be taken as a rationale for withholding emerging clinical observations or biologically plausible concerns. Ensuring that clinicians and patients have access to evolving evidence, including rare but mechanistically credible adverse events, is consistent with the core principles of medical ethics and reinforces public trust by clearly delineating what is known, what remains unresolved, and what work is ongoing. Informed consent must adapt as new data accumulate, even when causal questions remain open, and doing so does not assume bias to what future investigation will ultimately show.

CONCLUSIONS

The collective world-wide evidence from 2020–2025 underscores a biologically plausible connection between COVID-19 vaccination and cancer. The recurring clinical findings documented across many reports of de-novo cancer onset, rapid tumor progression, viral reactivation, and reawakening of dormant disease, highlight critical gaps in knowledge and understanding of how large-scale immune changes produced by the vaccine interacts with cancer biology.

Both SARS-CoV-2 infection and COVID-19 vaccination engage overlapping biological pathways that could, in principle, influence cancer risk, yet they differ in mechanism, magnitude, biodistribution, and duration of their effects. Shared mechanisms include activation of the innate immune system, robust interferon signaling, cytokine induction, oxidative stress, and transient disruption of immune-cell homeostasis. These changes can theoretically expose latent malignancies, promote clonal expansion of preexisting mutant cells, or create microenvironmental contexts permissive to tumor progression.

In addition, both infection and vaccination induce spike protein expression, which interacts with ACE2-expressing tissues and can trigger endothelial activation, inflammation, and cellular stress pathways implicated in oncogenic signaling. Both can also lead to prolonged inflammatory and tissue-injury states, all of which could contribute to genomic instability, epigenetic remodeling, and chronic immune dysregulation.

However, unique mechanisms distinguish COVID-19 vaccination from natural infection. Vaccination involves widespread biodistribution, intracellular uptake and persistence of modified nucleic acid templates that drive synthesis of an unnatural Spike protein both at the injection site but also throughout the body. The presence of the residual or fragmented DNA combined with the LNP-mediated delivery to immune and non-immune tissues, and sustained spike expression for month to years represent vaccine-specific factors that could theoretically promote insertional mutagenesis, perturb immune surveillance, or accelerate growth of preexisting malignant clones. As such there is much to be learned from human tissue and blood samples as well as autopsies to better understand the interplay between COVID infection, vaccination and cancer mechanisms.

To this end, Spike protein presence and persistence along with the biological effects that are cell autonomous or that depend on host immune interactions need to be studied to establish connections to cancer initiation and progression. Accordingly, we propose that tumors arising after documented SARS-CoV-2 infection or following COVID-19 vaccination be evaluated using a standardized immunohistochemical classification framework.

At minimum, this should include assessment of viral antigen expression patterns by IHC. Spike-positive/nucleocapsid-positive, spike-positive/nucleocapsid-negative, and spike-negative/nucleocapsid-negative phenotypes should be defined. This assessment should be integrated with detailed characterization of proliferative activity (e.g., Ki-67), cell-death and DNA-damage response markers, tumor-suppressor and oncogene pathway signatures, and the immune tumor microenvironment.

Implementing this type of reporting across clinical pathology and autopsy evaluations would allow more

precise discrimination among tumors potentially driven by infection, by vaccine-related antigen expression, or by unrelated oncogenic processes, and would enable aggregation of comparable cases across institutions. Establishing uniform criteria of this kind is essential for building a coherent evidence base, supporting mechanistic research, and ultimately determining whether observed associations reflect coincidence, unmasking of latent disease, immune perturbation, or true causal relationships.

Establishing causality between SARS-CoV-2 infection, COVID-19 vaccination, and cancer requires a level of evidence far beyond temporal association. In oncology, causation is never determined by a single observation or study; it emerges only when multiple, independent lines of evidence converge over time. This includes mechanistic data (such as genomic integration analyses, clonal evolution trajectories, immune-profiling, and epigenetic changes), pathology-based findings (including autopsies with molecular characterization), experimental models that accurately reflect human tissue biology (organoids, humanized systems, long-read sequencing of exposed tissues), and population-level epidemiologic studies capable of detecting small but meaningful signals against background incidence. Only by integrating these approaches can we distinguish coincidence from unmasking of latent disease, expansion of preexisting malignant clones, or true *de novo* oncogenesis. Importantly, the need for rigorous evidence should not be used to dismiss emerging patterns.

Transparent discussion of biologically plausible mechanisms and surveillance strategies is essential to determine if this temporal association is causally linked. Current reliable epidemiologic data is lacking to provide evidence that vaccination does not increase population-level cancer incidence. Peer-reviewed literature is not completely or easily indexed. Establishing a framework for post-vaccination cancer surveillance, could help detect rare adverse patterns early and enable mechanistic follow-up without compromising public confidence. The goal of this review is not to estimate population-level cancer risk but to provide a structured synthesis of the existing peer-reviewed literature, identify recurring clinical and biological themes, and delineate critical gaps that require rigorous epidemiologic and mechanistic follow-up. This will enable a better understanding of the full spectrum of immune responses to inform safer immunization strategies and illuminate previously underappreciated links between immunity and cancer biology.

The scientific imperative moving forward should be a coordinated framework combining longitudinal surveillance and mechanistic experimentation to allow us to distinguish coincidence from causality and to refine future vaccine platforms accordingly. In doing so, we stand to gain not only a clearer understanding of vaccine safety, but also a deeper insight into the fundamental links between immunity, infection, and cancer emergence.

AUTHOR CONTRIBUTIONS

C.K. and W.E-D. conceived the review, conducted the literature search, data extraction, and synthesis, drafted and approved the final version of the manuscript.

ACKNOWLEDGMENTS

W.S.E-D. is an American Cancer Society Research Professor and is supported by the Menoff Family University Professorship at Brown University.

CK is the Director of the Tufts University Convergence Laboratory of Biomedical, Physical, and Engineering Sciences.

CONFLICTS OF INTEREST

W.S.E-D., co-Editor-in-Chief of *Oncotarget*, was not involved in the review of this manuscript or the decision to accept it.

EDITORIAL NOTE

The Editor-in-Chief, Dr. Wafik S. El-Deiry, was not involved in the peer-review process or the decision-making for this paper. Dr. El-Deiry shared the submitted manuscript with National Cancer Institute (NCI) Director Anthony Letai by email electronically on December 12, 2025.

REFERENCES

1. Saha A, Ghosh Roy S, Dwivedi R, Tripathi P, Kumar K, Nambiar SM, Pathak R. Beyond the Pandemic Era: Recent Advances and Efficacy of SARS-CoV-2 Vaccines Against Emerging Variants of Concern. *Vaccines* (Basel). 2025; 13:424. <https://doi.org/10.3390/vaccines13040424>. [PubMed]
2. Chakraborty C, Lo YH, Bhattacharya M, Das A, Wen ZH. Looking beyond the origin of SARS-CoV-2: Significant strategic aspects during the five-year journey of COVID-19 vaccine development. *Mol Ther Nucleic Acids*. 2025; 36:102527. <https://doi.org/10.1016/j.omtn.2025.102527>. [PubMed]
3. Kumar A, Ahmad F, Sah BK, Aljabali AAA, Mishra Y, Mishra V. Advancements in viral vaccine development: from traditional to modern approaches. *Explor Immunol*. 2025; 5:1003203. <https://doi.org/10.37349/ ei.2025.1003203>.
4. Papadatou I, Michos A. Advances in Biotechnology and the Development of Novel Human Vaccines. *Vaccines* (Basel). 2025; 13:989. <https://doi.org/10.3390/vaccines13090989>. [PubMed]
5. Yang D, Tian J, Shen C, Li Q. An overview and single-arm meta-analysis of immune-mediated adverse events following COVID-19 vaccination. *Front Pharmacol*. 2024; 15:1308768. <https://doi.org/10.3389/fphar.2024.1308768>. [PubMed]

6. Buoninfante A, Andeweg A, Genov G, Cavalieri M. Myocarditis associated with COVID-19 vaccination. *NPJ Vaccines*. 2024; 9:122. <https://doi.org/10.1038/s41541-024-00893-1>. [PubMed]
7. Tayebi A, Samimisedeh P, Jafari Afshar E, Mahmoudnia S, Milan N, Ayati A, Madady A, Rastad H. Neuromuscular diseases associated with COVID-19 vaccines: a systematic review and pooled analysis of 258 patients. *BMC Neurol*. 2023; 23:437. <https://doi.org/10.1186/s12883-023-03486-y>. [PubMed]
8. Kim HJ, Kim MH, Choi MG, Chun EM. 1-year risks of cancers associated with COVID-19 vaccination: a large population-based cohort study in South Korea. *Biomark Res*. 2025; 13:114. <https://doi.org/10.1186/s40364-025-00831-w>. [PubMed]
9. Naffaa MM, Al-Ewaidat OA. The viral oncogenesis of COVID-19 and its impact on cancer progression, long-term risks, treatment complexities, and research strategies. *Explor Med*. 2025; 6:1001314. <https://doi.org/10.37349/emed.2025.1001314>.
10. Soleimani P, Yadollahifarsani S, Motieian M, Moghadam MSC, Alimohammadi S, Khayyat A, Pour MAE, Sina Neshat S, Marashi NA, Mahmoodnia L, Masomi R. Cancer recurrence or aggravation following COVID-19 vaccination. *J Nephropharmacol*. 2023; 12:e10593. <https://doi.org/10.34172/npj.2023.10593>.
11. Shi Y, Shi M, Wang Y, You J. Progress and prospects of mRNA-based drugs in pre-clinical and clinical applications. *Signal Transduct Target Ther*. 2024; 9:322. <https://doi.org/10.1038/s41392-024-02002-z>. [PubMed]
12. Comirnaty, t. P.-B. C.-m. v. <https://www.fda.gov/media/151707/download>.
13. Spikevax, t. M. C.-m. v. <https://www.fda.gov/media/155675/download>.
14. Mnexspike t. M. C.-l. d. m. v. <https://www.fda.gov/files/vaccines%20blood%20%26%20biologics/published/Package-Insert-MNEXSPIKE.pdf>.
15. Nuvaxovid t. N. C.-p.-b. v. <https://www.fda.gov/files/vaccines%20blood%20%26%20biologics/published/Package-Insert-and-Patient-Package-Insert-NUVAXOVID.pdf>.
16. Sung H, Jiang C, Bandi P, Minihan A, Fidler-Benaoudia M, Islami F, Siegel RL, Jemal A. Differences in cancer rates among adults born between 1920 and 1990 in the USA: an analysis of population-based cancer registry data. *Lancet Public Health*. 2024; 9:e583–93. [https://doi.org/10.1016/S2468-2667\(24\)00156-7](https://doi.org/10.1016/S2468-2667(24)00156-7). [PubMed]
17. Koh B, Tan DJH, Ng CH, Fu CE, Lim WH, Zeng RW, Yong JN, Koh JH, Syn N, Meng W, Wijarnpreecha K, Liu K, Chong CS, et al. Patterns in Cancer Incidence Among People Younger Than 50 Years in the US, 2010 to 2019. *JAMA Netw Open*. 2023; 6:e2328171. <https://doi.org/10.1001/jamanetworkopen.2023.28171>. [PubMed]
18. Siegel RL, Kratzer TB, Giaquinto AN, Sung H, Jemal A. Cancer statistics, 2025. *CA Cancer J Clin*. 2025; 75:10–45. <https://doi.org/10.3322/caac.21871>. [PubMed]
19. Grippin AJ, Marconi C, Copling S, Li N, Braun C, Woody C, Young E, Gupta P, Wang M, Wu A, Jeong SD, Soni D, Weidert F, et al, and D3CODE Team. SARS-CoV-2 mRNA vaccines sensitize tumours to immune checkpoint blockade. *Nature*. 2025; 647:488–97. <https://doi.org/10.1038/s41586-025-09655-y>. [PubMed]
20. Acuti Martellucci C, Capodici A, Soldato G, Fiore M, Zauli E, Carota R, De Benedictis M, Di Marco G, Di Luzio R, Flacco ME, Manzoli L. COVID-19 vaccination, all-cause mortality, and hospitalization for cancer: 30-month cohort study in an Italian province. *EXCLI J*. 2025; 24:690–707. <https://doi.org/10.17179/excli2025-8400>. [PubMed]
21. Bae E, Bae S, Vaysblat M, Abdelwahed M, Sarkar K, Bae S. Development of High-Grade Sarcoma After Second Dose of Moderna Vaccine. *Cureus*. 2023; 15:e37612. <https://doi.org/10.7759/cureus.37612>. [PubMed]
22. Quintero D, Patel N, Harris G, Maristany A, Alani A, Rosenberg AE, Conway SA, Jose J. COVID-19 vaccine-associated granulomatous mass mimicking a sarcoma: a case report. *Radiol Case Rep*. 2022; 17:2775–78. <https://doi.org/10.1016/j.radcr.2022.05.035>. [PubMed]
23. Li YH, Lin YT, Chuang SH, Yang HJ. Kaposi Sarcoma as a Possible Cutaneous Adverse Effect of ChAdOx1 nCov-19 Vaccine: A Case Report. *Vaccines (Basel)*. 2024; 12:1168. <https://doi.org/10.3390/vaccines12101168>. [PubMed]
24. White E, Fazio N, Tourmouzis K, Ryu S, Finger PT, Sasoon J, Keresztes R, Chou T, Kaplowitz K, Honkanen R. Unilateral conjunctival Classic Kaposi Sarcoma following a COVID 19 booster. *Am J Ophthalmol Case Rep*. 2023; 34:101986. <https://doi.org/10.1016/j.ajoc.2023.101986>. [PubMed]
25. Martínez-Ortega JI, Ramirez Cibrian AG, Martinez-Jaramillo E, García Silva MDC. Sporadic Kaposi Sarcoma Following a COVID-19 Vaccine: Mere Coincidence or Something More? *Cureus*. 2024; 16:e53925. <https://doi.org/10.7759/cureus.53925>. [PubMed]
26. Cavanna L, Grassi SO, Ruffini L, Michieletti E, Carella E, Palli D, Zangrandi A, Inzerilli N, Bernuzzi P, Di Nunzio C, Citterio C. Non-Hodgkin Lymphoma Developed Shortly after mRNA COVID-19 Vaccination: Report of a Case and Review of the Literature. *Medicina (Kaunas)*. 2023; 59:157. <https://doi.org/10.3390/medicina59010157>. [PubMed]
27. Olszewska B, Zaryckańska A, Nowicki RJ, Sokolowska-Wojdylo M. Rare COVID-19 vaccine side effects got lost in the shuffle. Primary cutaneous lymphomas following COVID-19 vaccination: a systematic review. *Front Med (Lausanne)*. 2024; 11:1325478. <https://doi.org/10.3389/fmed.2024.1325478>. [PubMed]
28. Sekizawa A, Hashimoto K, Kobayashi S, Kozono S, Kobayashi T, Kawamura Y, Kimata M, Fujita N, Ono Y, Obuchi Y, Tanaka Y. Rapid progression of marginal zone B-cell lymphoma after COVID-19 vaccination (BNT162b2): A case report. *Front Med (Lausanne)*. 2022; 9:963393. <https://doi.org/10.3389/fmed.2022.963393>. [PubMed]

29. Tosun A, Tasdemir MN. Glioblastoma misdiagnosed as acute disseminated encephalomyelitis following coronavirus disease vaccination. *Rev Soc Bras Med Trop.* 2022; 55:e0400–2022. <https://doi.org/10.1590/0037-8682-0400-2022>. [PubMed]

30. Akkus E, Karaoglan B, Akyol C, Ünal AE, Kuzu MA, Savaş B, Utkan G. Types and Rates of COVID-19 Vaccination in Patients With Newly Diagnosed Microsatellite Stable and Instable Non-Metastatic Colon Cancer. *Cureus.* 2024; 16:e61780. <https://doi.org/10.7759/cureus.61780>. [PubMed]

31. Kim JH, Oh EH, Han DS. Polyposis of gastrointestinal tract after COVID-19 mRNA vaccination: a report of two cases. *Clin Endosc.* 2024; 57:402–6. <https://doi.org/10.5946/ce.2023.268>. [PubMed]

32. Abue M, Mochizuki M, Shibuya-Takahashi R, Ota K, Wakui Y, Iwai W, Kusaka J, Saito M, Suzuki S, Sato I, Tamai K. Repeated COVID-19 Vaccination as a Poor Prognostic Factor in Pancreatic Cancer: A Retrospective, Single-Center Cohort Study. *Cancers (Basel).* 2025; 17:2006. <https://doi.org/10.3390/cancers17122006>. [PubMed]

33. Sasa S, Inoue H, Inui T, Miyamoto N, Aoyama M, Okumura K, Toba H, Yoshida T, Tezuka M, Hirose C, Saijo Y, Uehara H, Izumori A, et al. Axillary lymphangioma that developed following COVID-19 vaccination: a case report. *Surg Case Rep.* 2022; 8:131. <https://doi.org/10.1186/s40792-022-01488-5>. [PubMed]

34. Ang SY, Huang YF, Chang CT. Ph-Positive B-Cell Acute Lymphoblastic Leukemia Occurring after Receipt of Bivalent SARS-CoV-2 mRNA Vaccine Booster: A Case Report. *Medicina (Kaunas).* 2023; 59:627. <https://doi.org/10.3390/medicina59030627>. [PubMed]

35. Yanagida E, Kubota A, Miyoshi H, Ohshima K, Kawakita T, Murayama T. The case of T-ALL presenting with NK phenotype after COVID-19 vaccination. *Pathol Res Pract.* 2023; 242:154310. <https://doi.org/10.1016/j.prp.2023.154310>. [PubMed]

36. Sano S. A case of metastatic breast carcinoma to the skin expressing SARS-CoV-2 spike protein possibly derived from mRNA vaccine. *J Dermatol Sci.* 2025; 120:71–73. <https://doi.org/10.1016/j.jdermsci.2025.09.007>. [PubMed]

37. Mizutani M, Mitsui H, Amano T, Ogawa Y, Deguchi N, Shimada S, Miwa A, Kawamura T, Ogido Y. Two cases of axillary lymphadenopathy diagnosed as diffuse large B-cell lymphoma developed shortly after BNT162b2 COVID-19 vaccination. *J Eur Acad Dermatol Venereol.* 2022; 36:e613–15. <https://doi.org/10.1111/jdv.18136>. [PubMed]

38. Peacock JG, Banks EA, McWhorter N. ¹⁸F-Fluciclovine-Avid Axillary Lymph Nodes After COVID-19 Vaccination on PET/CT for Suspected Recurrence of Prostate Cancer. *J Nucl Med Technol.* 2022; 50:73–74. <https://doi.org/10.2967/jnmt.121.263001>. [PubMed]

39. Revenga-Porcel L, Peñate Y, Granados-Pacheco F. Anaplastic large cell lymphoma at the SARS-CoV2 vaccine injection site. *J Eur Acad Dermatol Venereol.* 2023; 37:e32–34. <https://doi.org/10.1111/jdv.18615>. [PubMed]

40. Hsieh CY, Yu SC, Lee JA, Tsai TF, Nodal T-Follicular Helper Cell Lymphoma, Angioimmunoblastic-Type, Diagnosed in a Patient with Psoriasis Following COVID-19 Vaccination under Adalimumab Treatment: A Causal Association? *Indian J Dermatol.* 2024; 69:264–67. https://doi.org/10.4103/ijd.ijd_93_23. [PubMed]

41. Tachita T, Takahata T, Yamashita S, Ebina T, Kamata K, Yamagata K, Tamai Y, Sakuraba H. Newly diagnosed extranodal NK/T-cell lymphoma, nasal type, at the injected left arm after BNT162b2 mRNA COVID-19 vaccination. *Int J Hematol.* 2023; 118:503–507. <https://doi.org/10.1007/s12185-023-03607-w>. [PubMed]

42. Tanaka A, Kawaguchi T, Imadome KI, Hara S. [Epstein-Barr virus-associated lymphoproliferative disorders after BNT162b2 mRNA COVID-19 vaccination]. *Rinsho Ketsueki.* 2023; 64:277–82. <https://doi.org/10.11406/rinketsu.64.277>. [PubMed]

43. Gambichler T, Boms S, Hessam S, Tischoff I, Tannapfel A, Lüttringhaus T, Beckman J, Stranzenbach R. Primary cutaneous anaplastic large-cell lymphoma with marked spontaneous regression of organ manifestation after SARS-CoV-2 vaccination. *Br J Dermatol.* 2021; 185:1259–62. <https://doi.org/10.1111/bjd.20630>. [PubMed]

44. Goldman S, Bron D, Tousseyen T, Vieras I, Dewispelaere L, Heimann P, Cogan E, Goldman M. Rapid Progression of Angioimmunoblastic T Cell Lymphoma Following BNT162b2 mRNA Vaccine Booster Shot: A Case Report. *Front Med (Lausanne).* 2021; 8:798095. <https://doi.org/10.3389/fmed.2021.798095>. [PubMed]

45. Dyagil I, Martina Z, Movchan A, Abramenco I, Chumak A, Bazyka D. Rapid progression of angioimmunoblastic T cell lymphoma after Covid-19 vaccination (Chadox1-S): a case report. *J Clin Case Rep Med Image Health Sci.* 2023; 5:2. <https://dx.doi.org/10.5592/JCRMHS.2023.05.001201>.

46. Panou E, Nikolaou V, Marinos L, Kallambou S, Sidiropoulou P, Gerochristou M, Stratigos A. Recurrence of cutaneous T-cell lymphoma post viral vector COVID-19 vaccination. *J Eur Acad Dermatol Venereol.* 2022; 36:e91–93. <https://doi.org/10.1111/jdv.17736>. [PubMed]

47. Kreher MA, Ahn J, Werbel T, Motaparthi K. Subcutaneous panniculitis-like T-cell lymphoma after COVID-19 vaccination. *JAAD Case Rep.* 2022; 28:18–20. <https://doi.org/10.1016/j.jdcr.2022.08.006>. [PubMed]

48. Ukishima S, Miyagami T, Arikawa M, Kushiro S, Takaku T, Naito T. Subcutaneous panniculitis-like T-cell lymphoma post-mRNA-1273 COVID-19 vaccination. *Clin Case Rep.* 2023; 11:e7143. <https://doi.org/10.1002/ccr3.7143>. [PubMed]

49. Veeraballi S, Patel A, Are G, Ramahi A, Chittamuri S, Shaaban H. A Case of Chronic Myelomonocytic Leukemia Unmasked After Receiving J&J COVID-19 Vaccine. *Cureus.* 2022; 14:e26070. <https://doi.org/10.7759/cureus.26070>. [PubMed]

50. Plüß M, Mitteldorf C, Szuszies CJ, Tampe B. Case Report: Acquired Haemophilia A Following mRNA-1273 Booster Vaccination Against SARS-CoV-2 With

Concurrent Diagnosis of Pleomorphic Dermal Sarcoma. *Front Immunol.* 2022; 13:868133. <https://doi.org/10.3389/fimmu.2022.868133>. [PubMed]

51. Sumi T, Nagahisa Y, Matsuura K, Sekikawa M, Yamada Y, Nakata H, Chiba H. Lung squamous cell carcinoma with hemoptysis after vaccination with tozinameran (BNT162b2, Pfizer-BioNTech). *Thorac Cancer.* 2021; 12:3072–75. <https://doi.org/10.1111/1759-7714.14179>. [PubMed]

52. Eens S, Van Hecke M, Favere K, Toussaint T, Guns PJ, Roskams T, Heidbuchel H. B-cell lymphoblastic lymphoma following intravenous BNT162b2 mRNA booster in a BALB/c mouse: A case report. *Front Oncol.* 2023; 13:1158124. <https://doi.org/10.3389/fonc.2023.1158124>. [PubMed]

53. Cui Z, Cheng J. A systematic review of lymphoma secondary to COVID-19 vaccination. *Authorea.* 2024. <https://doi.org/10.22541/au.172543425.59165134/v1>.

54. Barnett C, Mehta N, Towne WS, Babagbemi K, Sales RM. Metastatic melanoma in the breast and axilla: A case report. *Clin Imaging.* 2022; 85:78–82. <https://doi.org/10.1016/j.clinimag.2022.02.014>. [PubMed]

55. Gullotti DM, Lipson EJ, Fishman EK, Rowe SP. Acute axillary lymphadenopathy detected shortly after COVID-19 vaccination found to be due to newly diagnosed metastatic melanoma. *Radiol Case Rep.* 2022; 17:878–80. <https://doi.org/10.1016/j.radcr.2021.12.002>. [PubMed]

56. Wagle AM, Wu BC, Gopal L, Sundar G. Necrosis of uveal melanoma post-COVID-19 vaccination. *Indian J Ophthalmol.* 2022; 70:1837–40. https://doi.org/10.4103/ijo.IJO_3040_21. [PubMed]

57. Leis AA, Montesi AP, Khan SM, Montesi M. Case Report: Malignant Melanoma Associated With COVID-19: A Coincidence or a Clue? *Front Med (Lausanne).* 2022; 9:845558. <https://doi.org/10.3389/fmed.2022.845558>. [PubMed]

58. Avram A, Scurtu LG, Costache M, Simionescu O. A Disguising Fast-Growing Metachronous Melanoma and COVID-19. *Cureus.* 2023; 15:e36108. <https://doi.org/10.7759/cureus.36108>. [PubMed]

59. Sprenger F, Sanches DP, Bacarin JV, de Almeida Teixeira BC. Prevertebral Inflammatory Myofibroblastic Tumor Following COVID Vaccine Booster Dose. *Indian J Otolaryngol Head Neck Surg.* 2023; 75:1–4. <https://doi.org/10.1007/s12070-023-03718-0>. [PubMed]

60. Sugashawa S, Kimura T, Bae Y, Kumasaka T, Ichi S. Two Cases of Rare Intratumoral Hemorrhage Following COVID-19 Vaccination. *Cureus.* 2022; 14:e32400. <https://doi.org/10.7759/cureus.32400>. [PubMed]

61. Gable AD, Hughes SM, Miller RJ. A 24-Year-Old Man With Hemoptysis Found to Have a Chest Mass and Contralateral Axillary Lymphadenopathy. *Chest.* 2021; 160:e289–93. <https://doi.org/10.1016/j.chest.2021.04.055>. [PubMed]

62. Xu W, Nian W. A rare adverse effects of COVID-19 vaccine in a patient with a latent tumor: A case report and literature review. *Front Oncol.* 2023; 13:1269735. <https://doi.org/10.3389/fonc.2023.1269735>. [PubMed]

63. Farhat M, Zouein J, Abou Khater J, Sarkis AS, Helou J. A Case of Rapid Transformation of a Nail Matrix Nevi to Melanoma After Messenger RNA COVID-19 Vaccine: A Cause or a Coincidence? *Cureus.* 2024; 16:e76312. <https://doi.org/10.7759/cureus.76312>. [PubMed]

64. Bogomolets O, Rojczyk E, Hryshchenko R, Bogomolets C, Berezhkin O. Covid-19, leukemia, and secondary malignancies of the skin - is there a connection: a case report and literature analysis. *Front Oncol.* 2023; 13:1265479. <https://doi.org/10.3389/fonc.2023.1265479>. [PubMed]

65. Abbas AM, Caicedo ML, Damron TA. Right Biceps Pseudo-Tumor from COVID-19 Vaccination. *Vaccines (Basel).* 2024; 12:160. <https://doi.org/10.3390/vaccines12020160>. [PubMed]

66. Iliadis AS, Koletsas T, Vounotrypidis P, Fassas A, Apostolidis A, Apostolidis S, Fotiadou A, Hytioglou P. Inflammatory Pseudotumor of the Spleen in a Patient With Psoriatic Arthritis: A Diagnostic Challenge in the COVID-19 Era. *Cureus.* 2024; 16:e73121. <https://doi.org/10.7759/cureus.73121>. [PubMed]

67. Evangelista FB. Dermatofibrosarcoma Protuberans following COVID-19 and Influenza vaccination: A Case Report. *Int J Med Rev Case Rep.* 2025; 25:1–6. <https://www.researchgate.net/publication/390665272>
Dermatofibrosarcoma_Protuberans_following_COVID-19_and_Influenza_vaccination_A_Case_Report

68. Yilmaz A, Goker B, Gedikoglu MG, Ayvaz M, Tokgozoglu AM. Primary Cutaneous Adenoid Cystic Carcinoma in a Rare Location With an Immune Response to a BNT162b2 Vaccine: A Case Report. *JBJS Case Connect.* 2024; 14. <https://doi.org/10.2106/JBJS.CC.23.00499>. [PubMed]

69. Rashid B, Baig I, Feroze MD, Chaudhry M, Hassan MSZ. Incidental Atrial Myxoma in the Shadow of COVID-19: Coincidence or an Emerging Pattern? *Cureus.* 2025; 17:e93617. <https://doi.org/10.7759/cureus.93617>. [PubMed]

70. Kawamoto H, Saito-Sasaki N, Sawada Y. Recurrent Merkel Cell Carcinoma Following COVID-19 Treatment. *Cureus.* 2024; 16:e64863. <https://doi.org/10.7759/cureus.64863>. [PubMed]

71. Bharathidasan K, Tran V, Ghafouri SR, Rehman S, Brandi L. Metastatic prostatic adenocarcinoma presenting as generalized lymphadenopathy unmasked by a COVID booster vaccine. *Clin Case Rep.* 2023; 11:e8278. <https://doi.org/10.1002/ccr3.8278>. [PubMed]

72. Hu H, Wang C, Tao R, Liu B, Peng D, Chen Y, Zhang W. Evidences of neurological injury caused by COVID-19 from glioma tissues and glioma organoids. *CNS Neurosci Ther.* 2024; 30:e14822. <https://doi.org/10.1111/cns.14822>. [PubMed]

73. Gregory TA, Knight SR, Aaroe AE, Highsmith KN, Janatpour ZC, O'Brien BJ, Majd NK, Loghin ME, Patel

CB, Weathers SP, Puduvali VK, Kamiya-Matsuoka C. Accelerated tumor progression after COVID-19 infection in patients with glioblastoma: A retrospective case-control study. *Neurooncol Pract.* 2024; 11:475–83. <https://doi.org/10.1093/nop/npae029>. [PubMed]

74. Chia SB, Johnson BJ, Hu J, Valenca-Pereira F, Chadeau-Hyam M, Guntoro F, Montgomery H, Boorgula MP, Sreekanth V, Goodspeed A, Davenport B, De Dominicis M, Zaberezhnyy V, et al. Respiratory viral infections awaken metastatic breast cancer cells in lungs. *Nature.* 2025; 645:496–506. <https://doi.org/10.1038/s41586-025-09332-0>. [PubMed]

75. Khatri JK, Tahboub I, Anwar K, Masoudi M, Graffeo V, Jamil MO. Diagnosis of Angioimmunoblastic T Cell Lymphoma After Receiving First Dose of Pfizer/BioNTech (BNT162b2) Vaccine: A Case Report. *J Investig Med High Impact Case Rep.* 2024; 12:23247096241231645. <https://doi.org/10.1177/23247096241231645>. [PubMed]

76. Obodo OI, Ocheni S, Obodo CO, Duru AN, Okoye HC, Nonyelu CE, Anigbogu IO, Nwaghwa TU, Madu AJ. Lymphoid Neoplasms and COVID-19 Vaccination. *Oncol Adv.* 2025; 3:e00005. <https://doi.org/10.14218/OnA.2025.00005>.

77. Ueda Y, Sakai T, Yamada K, Arita K, Ishige Y, Hoshi D, Yanagisawa H, Iwao-Kawanami H, Kawanami T, Mizuta S, Fukushima T, Yamada S, Yachie A, Masaki Y. Fatal hemophagocytic lymphohistiocytosis with intravascular large B-cell lymphoma following coronavirus disease 2019 vaccination in a patient with systemic lupus erythematosus: an intertwined case. *Immunol Med.* 2024; 47:192–99. <https://doi.org/10.1080/25785826.2024.2338594>. [PubMed]

78. Embaby A, Ibrahim A, Elsheikh H, AlMalki M, Kashif M, Althomali M, Babateen A. Diffuse Large B-Cell Lymphoma Developed Shortly After mRNA COVID-19 Vaccination in a Patient with Sickle Cell Disease: A Chicken-and-Egg Problem. *Bratisl Lek Listy.* 2025; 126:1997–83. <https://link.springer.com/article/10.1007/s44411-025-00174-w>.

79. Avallone G, Marone CA, Conforti C, Fava P, Gargiulo L, Marzano AV, Massone C, Mastorino L, Paradisi A, Pileri A, Quaglino P, Rizzo N, Riberio S, et al. Real-world data on primary cutaneous lymphoproliferative disorders following SARS-CoV-2 vaccination: A multicentre experience from tertiary referral hospitals. *J Eur Acad Dermatol Venereol.* 2023; 37:e451–55. <https://doi.org/10.1111/jdv.18806>. [PubMed]

80. Gordon ER, Kwinta BD, Schreidah CM, Fahmy LM, Adeuyan O, Queen D, Trager MH, Magro CM, Geskin LJ. Cutaneous lymphoproliferative disorders after COVID-19 vaccination: clinical presentation, histopathology, and outcomes. *Leuk Lymphoma.* 2024; 65:48–54. <https://doi.org/10.1080/10428194.2023.2270766>. [PubMed]

81. Hobayan CG, Chung CG. Indolent cutaneous lymphoma with gamma/delta expression after COVID-19 vaccination. *JAAD Case Rep.* 2023; 32:74–76. <https://doi.org/10.1016/j.jdcr.2022.12.001>. [PubMed]

82. Moneva-Sakelarieva M, Kobakova Y, Konstantinov S, Semerdzhieva NE, Ivanova SA, Atanasova VP, Atanasov PY, Becheva MSV. Lymphoma with skin localization and COVID 19 – a clinical case and review of literature. *Pharmacia.* 2024; 71:1–7. <https://doi.org/10.3897/pharmacia.71.e135467>.

83. Montoya VHG, Cardona LG, Morales S, Ospina JA, Rueda X. SARSCOV-2 vaccine associated with primary cutaneous peripheral T cell lymphoma. *Eur J Cancer.* 2022. [https://doi.org/10.1016/S0959-8049\(22\)00617-7](https://doi.org/10.1016/S0959-8049(22)00617-7).

84. O Sullivan C, Zach F, Moser T, Pilz G, Harrer A, Trinka E, Enzinger C, Pfaff JAR, Wipfler P. Misinterpretation of glioblastoma as ADEM: potentially harmful consequences of over-diagnosis of COVID-19 vaccine-associated adverse events. *J Neurol.* 2022; 269:616–18. <https://doi.org/10.1007/s00415-021-10707-2>. [PubMed]

85. Russell SJ, Mabila SL. Non-Hodgkin lymphoma incidence in active component U.S. service members, 2017–2023. *MSMR.* 2025; 32:16–17. [PubMed]

86. Mitrofanova L, Makarov I, Goncharova E, Makarova T, Starshinova A, Kudlay D, Shlaykhto E. High Risk of Heart Tumors after COVID-19. *Life (Basel).* 2023; 13:2087. <https://doi.org/10.3390/life13102087>. [PubMed]

87. Zeyen T, Friker LL, Paech D, Schaefer N, Weller J, Zschernack V, Layer JP, Schneider M, Potthoff AL, Bernhardt M, Sanders C, Kristiansen G, Hoelzel M, et al. Transient MRI changes and neurological deterioration in glioblastoma upon SARS-CoV-2 infection. *J Cancer Res Clin Oncol.* 2024; 150:437. <https://doi.org/10.1007/s00432-024-05963-4>. [PubMed]

88. Bradford PT, Devesa SS, Anderson WF, Toro JR. Cutaneous lymphoma incidence patterns in the United States: a population-based study of 3884 cases. *Blood.* 2009; 113:5064–73. <https://doi.org/10.1182/blood-2008-10-184168>. [PubMed]

89. Molyneux EM, Rochford R, Griffin B, Newton R, Jackson G, Menon G, Harrison CJ, Israels T, Bailey S. Burkitt's lymphoma. *Lancet.* 2012; 379:1234–44. [https://doi.org/10.1016/S0140-6736\(11\)61177-X](https://doi.org/10.1016/S0140-6736(11)61177-X). [PubMed]

90. Stoyanov GS, Lyutfi E, Georgiev R, Dzhenkov DL, Kaprelyan A. The Rapid Development of Glioblastoma: A Report of Two Cases. *Cureus.* 2022; 14:e26319. <https://doi.org/10.7759/cureus.26319>. [PubMed]

91. Stein WD, Figg WD, Dahut W, Stein AD, Hoshen MB, Price D, Bates SE, Fojo T. Tumor growth rates derived from data for patients in a clinical trial correlate strongly with patient survival: a novel strategy for evaluation of clinical trial data. *Oncologist.* 2008; 13:1046–54. <https://doi.org/10.1634/theoncologist.2008-0075>. [PubMed]

92. He LN, Zhang X, Li H, Chen T, Chen C, Zhou Y, Lin Z, Du W, Fang W, Yang Y, Huang Y, Zhao H, Hong S, Zhang L. Pre-Treatment Tumor Growth Rate Predicts Clinical

Outcomes of Patients With Advanced Non-Small Cell Lung Cancer Undergoing Anti-PD-1/PD-L1 Therapy. *Front Oncol.* 2021; 10:621329. <https://doi.org/10.3389/fonc.2020.621329>. [PubMed]

93. Endara SA, Dávalos GA, Molina GA, Zavala AB, Ponton PM, Brito M, Nieto C, Ullauri VE. COVID-19 infection and cardiac angiosarcoma: a dangerous combination-a case report. *Cardiothorac Surg.* 2021; 29:5. <https://doi.org/10.1186/s43057-021-00042-7>. [PubMed]

94. Hu M, Wang B, Li J, Wu C. Editorial: The association between viral infection and human cancers. *Front Microbiol.* 2024; 15:1371581. <https://doi.org/10.3389/fmicb.2024.1371581>. [PubMed]

95. Ameya G, Birri DJ. The molecular mechanisms of virus-induced human cancers. *Microb Pathog.* 2023; 183:106292. <https://doi.org/10.1016/j.micpath.2023.106292>. [PubMed]

96. McLaughlin-Drubin ME, Munger K. Viruses associated with human cancer. *Biochim Biophys Acta.* 2008; 1782:127–50. <https://doi.org/10.1016/j.bbapap.2007.12.005>. [PubMed]

97. Moore PS, Chang Y. Why do viruses cause cancer? Highlights of the first century of human tumour virology. *Nat Rev Cancer.* 2010; 10:878–89. <https://doi.org/10.1038/nrc2961>. [PubMed]

98. Morais P, Adachi H, Yu YT. The Critical Contribution of Pseudouridine to mRNA COVID-19 Vaccines. *Front Cell Dev Biol.* 2021; 9:789427. <https://doi.org/10.3389/fcell.2021.789427>. [PubMed]

99. Bettini E, Locci M. SARS-CoV-2 mRNA Vaccines: Immunological Mechanism and Beyond. *Vaccines (Basel).* 2021; 9:147. <https://doi.org/10.3390/vaccines9020147>. [PubMed]

100. Boros LG, Kyriakopoulos AM, Brogna C, Piscopo M, McCullough PA, Seneff S. Long-lasting, biochemically modified mRNA, and its frameshifted recombinant spike proteins in human tissues and circulation after COVID-19 vaccination. *Pharmacol Res Perspect.* 2024; 12:e1218. <https://doi.org/10.1002/prp2.1218>. [PubMed]

101. Mulroney TE, Pöyry T, Yam-Puc JC, Rust M, Harvey RF, Kalmar L, Horner E, Booth L, Ferreira AP, Stoneley M, Sawarkar R, Mentzer AJ, Lilley KS, et al. N¹-methylpseudouridylation of mRNA causes +1 ribosomal frameshifting. *Nature.* 2024; 625:189–94. <https://doi.org/10.1038/s41586-023-06800-3>. [PubMed]

102. Jackson LA, Anderson EJ, Roushafael NG, Roberts PC, Makhene M, Coler RN, McCullough MP, Chappell JD, Denison MR, Stevens LJ, Pruijssers AJ, McDermott A, Flach B, et al, and mRNA-1273 Study Group. An mRNA Vaccine against SARS-CoV-2 - Preliminary Report. *N Engl J Med.* 2020; 383:1920–31. <https://doi.org/10.1056/NEJMoa2022483>. [PubMed]

103. Speicher DJ, Rose J, McKernan K. Quantification of residual plasmid DNA and SV40 promoter-enhancer sequences in Pfizer/BioNTech and Moderna modRNA COVID-19 vaccines from Ontario, Canada. *Autoimmunity.* 2025; 58:2551517. <https://doi.org/10.1080/08916934.2025.2551517>. [PubMed]

104. Raoult D. Confirmation of the presence of vaccine DNA in the Pfizer anti-COVID-19 vaccine. 2024. <https://hal.science/hal-04778576v1>.

105. Kaiser S, Kaiser S, Reis J, Marschalek R. Quantification of objective concentrations of DNA impurities in mRNA vaccines. *Vaccine.* 2025; 55:127022. <https://doi.org/10.1016/j.vaccine.2025.127022>. [PubMed]

106. Science, Public Health Policy, and the Law Volume: v5.2019-2024. December 2024. https://www.researchgate.net/publication/386414408_BioNTech_RNA-Based_COVID-19_Injections_Contain_Large_Amounts_Of_Residual_DNA_Including_An_SV40_PromoterEnhancer_Sequence.

107. König B, Kirchner JO. Methodological Considerations Regarding the Quantification of DNA Impurities in the COVID-19 mRNA Vaccine Comirnaty®. *Methods Protoc.* 2024; 7:41. <https://doi.org/10.3390/mps7030041>. [PubMed]

108. Wang TJ, Kim A, Kim K. A rapid detection method of replication-competent plasmid DNA from COVID-19 mRNA vaccines for quality control. *JHSS.* 2024; 8. https://www.researchgate.net/publication/393592852_A_rapid_detection_method_of_replication-competent_plasmid_DNA_from_COVID-19_mRNA_vaccines_for_quality_control.

109. Patterson BK, Yogendra R, Francisco EB, Guevara-Coto J, Long E, Pise A, Osgood E, Bream J, Kreimer M, Jeffers D, Beaty C, Vander Heide R, Mora-Rodríguez RA. Detection of S1 spike protein in CD16+ monocytes up to 245 days in SARS-CoV-2-negative post-COVID-19 vaccine syndrome (PCVS) individuals. *Hum Vaccin Immunother.* 2025; 21:2494934. <https://doi.org/10.1080/21645515.2025.2494934>. [PubMed]

110. Bhattacharjee S. Immunological and Antigenic Signatures Associated with Chronic Illnesses after COVID-19 Vaccination. *medRxiv.* 2025. <https://www.medrxiv.org/content/10.1101/2025.02.18.25322379v1>.

111. Halma M, Varon J. Breaking the silence: Recognizing post-vaccination syndrome. *Heliyon.* 2025; 11:e43478. <https://doi.org/https://doi.org/10.1016/j.heliyon.2025.e43478>.

112. Banoun H. mRNA: Vaccine or Gene Therapy? The Safety Regulatory Issues. *Int J Mol Sci.* 2023; 24:10514. <https://doi.org/10.3390/ijms241310514>. [PubMed]

113. <https://www.fda.gov/vaccines-blood-biologics/cellular-gene-therapy-products/what-gene-therapy>.

114. https://www.europarl.europa.eu/doceo/document/E-9-2024-000355_EN.html.

115. Azzarone B, Landolina N, Mariotti FR, Moretta L, Maggi E. Soluble SARS-CoV-2 Spike glycoprotein: considering some potential pathogenic effects. *Front Immunol.* 2025; 16:1616106. <https://doi.org/10.3389/fimmu.2025.1616106>. [PubMed]

116. Swank Z, Senussi Y, Alter G, Walt DR. Persistent circulating SARS-CoV-2 spike is associated with post-acute COVID-19 sequelae. *medrxiv*. 2022. <https://doi.org/10.1101/2022.06.14.22276401>.

117. Cabău G, Badii M, Mirea AM, Gaal OI, van Emst L, Popp RA, Crișan TO, Joosten LAB. Long-Lasting Enhanced Cytokine Responses Following SARS-CoV-2 BNT162b2 mRNA Vaccination. *Vaccines (Basel)*. 2024; 12:736. <https://doi.org/10.3390/vaccines12070736>. [PubMed]

118. Mung SM, Goh TL, Hughes M, Jude EB. Inflammatory Arthritis Associated with COVID-19 Vaccination. *Cureus*. 2023; 15:e35951. <https://doi.org/10.7759/cureus.35951>. [PubMed]

119. Jagtap K, Naveen R, Day J, Sen P, Vaidya B, Nune A, Nikiphorou E, Tan AL, Agarwal V, Saha S, Shinjo SK, Ziade N, Joshi M, et al, and COVAD Study Group. Flares in autoimmune rheumatic diseases in the post-COVID-19 vaccination period-a cross-sequential study based on COVAD surveys. *Rheumatology (Oxford)*. 2023; 62:3838–48. <https://doi.org/10.1093/rheumatology/kead144>. [PubMed]

120. Lim S, Yocum RR, Silver PA, Way JC. High spontaneous integration rates of end-modified linear DNAs upon mammalian cell transfection. *Sci Rep*. 2023; 13:6835. <https://doi.org/10.1038/s41598-023-33862-0>. [PubMed]

121. Tahtinen S, Tong AJ, Himmels P, Oh J, Paler-Martinez A, Kim L, Wichner S, Oei Y, McCarron MJ, Freund EC, Amir ZA, de la Cruz CC, Haley B, et al. IL-1 and IL-1ra are key regulators of the inflammatory response to RNA vaccines. *Nat Immunol*. 2022; 23:532–42. <https://doi.org/10.1038/s41590-022-01160-y>. [PubMed]

122. Rosati M, Terpos E, Homan P, Bergamaschi C, Karalioti S, Ntanasis-Stathopoulos I, Devasundaram S, Bear J, Burns R, Bagratuni T, Trougakos IP, Dimopoulos MA, Pavlakis GN, Felber BK. Rapid transient and longer-lasting innate cytokine changes associated with adaptive immunity after repeated SARS-CoV-2 BNT162b2 mRNA vaccinations. *Front Immunol*. 2023; 14:1292568. <https://doi.org/10.3389/fimmu.2023.1292568>. [PubMed]

123. Kim S, Jeon JH, Kim M, Lee Y, Hwang YH, Park M, Li CH, Lee T, Lee JA, Kim YM, Kim D, Lee H, Kim YJ, et al. Innate immune responses against mRNA vaccine promote cellular immunity through IFN- β at the injection site. *Nat Commun*. 2024; 15:7226. <https://doi.org/10.1038/s41467-024-51411-9>. [PubMed]

124. Faraj SS, Jalal PJ. IL1 β , IL-6, and TNF- α cytokines cooperate to modulate a complicated medical condition among COVID-19 patients: case-control study. *Ann Med Surg (Lond)*. 2023; 85:2291–97. <https://doi.org/10.1097/MS9.0000000000000679>. [PubMed]

125. Korzun T, Moses AS, Jozic A, Grigoriev V, Newton S, Kim J, Diba P, Sattler A, Levasseur PR, Le N, Singh P, Sharma KS, Goo YT, et al. Lipid Nanoparticles Elicit Reactogenicity and Sickness Behavior in Mice Via Toll-Like Receptor 4 and Myeloid Differentiation Protein 88 Axis. *ACS Nano*. 2024; 18:24842–59. <https://doi.org/10.1021/acsnano.4c05088>. [PubMed]

126. Zhang L, More KR, Ojha A, Jackson CB, Quinlan BD, Li H, He W, Farzan M, Pardi N, Choe H. Effect of mRNA-LNP components of two globally-marketed COVID-19 vaccines on efficacy and stability. *NPJ Vaccines*. 2023; 8:156. <https://doi.org/10.1038/s41541-023-00751-6>. [PubMed]

127. Nguyen HM, Alexander KE, Collinge M, Hickey JC, Lanz TA, Li J, Sheehan MJ, Newman LC, Thorn M. mRNA-LNPs induce immune activation and cytokine release in human whole blood assays across diverse health conditions. *Mol Ther*. 2025; 33:2872–85. <https://doi.org/10.1016/j.mtthe.2024.12.019>. [PubMed]

128. Amoako K, Mokhammad A, Malik A, Yesudasan S, Wheba A, Olagunju O, Gu SX, Yarovinsky T, Faustino EVS, Nguyen J, Hwa J. Enhancing nucleic acid delivery by the integration of artificial intelligence into lipid nanoparticle formulation. *Front Med Technol*. 2025; 7:1591119. <https://doi.org/10.3389/fmedt.2025.1591119>. [PubMed]

129. Pateev I, Seregina K, Ivanov R, Reshetnikov V. Biodistribution of RNA Vaccines and of Their Products: Evidence from Human and Animal Studies. *Biomedicines*. 2023; 12:59. <https://doi.org/10.3390/biomedicines12010059>. [PubMed]

130. Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. *J Immunol Res*. 2014; 2014:149185. <https://doi.org/10.1155/2014/149185>. [PubMed]

131. Manjili SH, Isbell M, Ghochaghi N, Perkinson T, Manjili MH. Multifaceted functions of chronic inflammation in regulating tumor dormancy and relapse. *Semin Cancer Biol*. 2022; 78:17–22. <https://doi.org/10.1016/j.semcan.2021.03.023>. [PubMed]

132. Albrengues J, Shields MA, Ng D, Park CG, Ambrico A, Poindexter ME, Upadhyay P, Uyeminami DL, Pommier A, Küttner V, Bružas E, Maiorino L, Bautista C, et al. Neutrophil extracellular traps produced during inflammation awaken dormant cancer cells in mice. *Science*. 2018; 361:eaao4227. <https://doi.org/10.1126/science.aa04227>. [PubMed]

133. Perego M, Tyurin VA, Tyurina YY, Yellets J, Nacarelli T, Lin C, Nefedova Y, Kossenkov A, Liu Q, Sreedhar S, Pass H, Roth J, Vogl T, et al. Reactivation of dormant tumor cells by modified lipids derived from stress-activated neutrophils. *Sci Transl Med*. 2020; 12:eabb5817. <https://doi.org/10.1126/scitranslmed.abb5817>. [PubMed]

134. Maier JA, Castiglioni S, Petrelli A, Cannatelli R, Ferretti F, Pellegrino G, Sarzi Puttini P, Fiorina P, Ardizzone S. Immune-Mediated Inflammatory Diseases and Cancer - a dangerous liaison. *Front Immunol*. 2024; 15:1436581. <https://doi.org/10.3389/fimmu.2024.1436581>. [PubMed]

135. Sausen DG, Poirier MC, Spiers LM, Smith EN. Mechanisms of T cell evasion by Epstein-Barr virus and implications for

tumor survival. *Front Immunol.* 2023; 14:1289313. <https://doi.org/10.3389/fimmu.2023.1289313>. [PubMed]

136. Aneja KK, Yuan Y. Reactivation and Lytic Replication of Kaposi's Sarcoma-Associated Herpesvirus: An Update. *Front Microbiol.* 2017; 8:613. <https://doi.org/10.3389/fmicb.2017.00613>. [PubMed]

137. Traylen CM, Patel HR, Fondaw W, Mahatme S, Williams JF, Walker LR, Dyson OF, Arce S, Akula SM. Virus reactivation: a panoramic view in human infections. *Future Virol.* 2011; 6:451–63. <https://doi.org/10.2217/fvl.11.21>. [PubMed]

138. Okan Cakir M, Kirca O, Gunduz S, Ozdogan M. Hyperprogression after immunotherapy: A comprehensive review. *J BUON.* 2019; 24:2232–41. [PubMed]

139. Shen P, Han L, Ba X, Qin K, Tu S. Hyperprogressive Disease in Cancers Treated With Immune Checkpoint Inhibitors. *Front Pharmacol.* 2021; 12:678409. <https://doi.org/10.3389/fphar.2021.678409>. [PubMed]

140. Adashek JJ, Kato S, Ferrara R, Lo Russo G, Kurzrock R. Hyperprogression and Immune Checkpoint Inhibitors: Hype or Progress? *Oncologist.* 2020; 25:94–98. <https://doi.org/10.1634/theoncologist.2019-0636>. [PubMed]

141. Brooks DG, Teyton L, Oldstone MB, McGavern DB. Intrinsic functional dysregulation of CD4 T cells occurs rapidly following persistent viral infection. *J Virol.* 2005; 79:10514–27. <https://doi.org/10.1128/JVI.79.16.10514-10527.2005>. [PubMed]

142. Zhou S, Ou R, Huang L, Price GE, Moskophidis D. Differential tissue-specific regulation of antiviral CD8+ T-cell immune responses during chronic viral infection. *J Virol.* 2004; 78:3578–600. <https://doi.org/10.1128/jvi.78.7.3578-3600.2004>. [PubMed]

143. Zhou Y, Liao X, Song X, He M, Xiao F, Jin X, Xie X, Zhang Z, Wang B, Zhou C, Kang Y, Zhang W. Severe Adaptive Immune Suppression May Be Why Patients With Severe COVID-19 Cannot Be Discharged From the ICU Even After Negative Viral Tests. *Front Immunol.* 2021; 12:755579. <https://doi.org/10.3389/fimmu.2021.755579>. [PubMed]

144. Hotchkiss RS, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol.* 2013; 13:862–74. <https://doi.org/10.1038/nri3552>. [PubMed]

145. Irrgang P, Gerling J, Kocher K, Lapuente D, Steininger P, Habenicht K, Wytopil M, Beileke S, Schäfer S, Zhong J, Ssebyatika G, Krey T, Falcone V, et al. Class switch toward noninflammatory, spike-specific IgG4 antibodies after repeated SARS-CoV-2 mRNA vaccination. *Sci Immunol.* 2023; 8:eade2798. <https://doi.org/10.1126/sciimmunol.ade2798>. [PubMed]

146. Shi P, Yu Y, Xie H, Yin X, Chen X, Zhao Y, Zhao H. Recent advances in regulatory immune cells: exploring the world beyond Tregs. *Front Immunol.* 2025; 16:1530301. <https://doi.org/10.3389/fimmu.2025.1530301>. [PubMed]

147. Facciabene A, Motz GT, Coukos G. T-regulatory cells: key players in tumor immune escape and angiogenesis. *Cancer Res.* 2012; 72:2162–71. <https://doi.org/10.1158/0008-5472.CAN-11-3687>. [PubMed]

148. Devaud C, John LB, Westwood JA, Darcy PK, Kershaw MH. Immune modulation of the tumor microenvironment for enhancing cancer immunotherapy. *Oncoimmunology.* 2013; 2:e25961. <https://doi.org/10.4161/onci.25961>. [PubMed]

149. Yang Y, Li S, To KKW, Zhu S, Wang F, Fu L. Tumor-associated macrophages remodel the suppressive tumor immune microenvironment and targeted therapy for immunotherapy. *J Exp Clin Cancer Res.* 2025; 44:145. <https://doi.org/10.1186/s13046-025-03377-9>. [PubMed]

150. Veleri S. Neurotropism of SARS-CoV-2 and neurological diseases of the central nervous system in COVID-19 patients. *Exp Brain Res.* 2022; 240:9–25. <https://doi.org/10.1007/s00221-021-06244-z>. [PubMed]

151. Erickson MA, Logsdon AF, Rhea EM, Hansen KM, Holden SJ, Banks WA, Smith JL, German C, Farr SA, Morley JE, Weaver RR, Hirsch AJ, Kovac A, et al. Blood-brain barrier penetration of non-replicating SARS-CoV-2 and S1 variants of concern induce neuroinflammation which is accentuated in a mouse model of Alzheimer's disease. *Brain Behav Immun.* 2023; 109:251–68. <https://doi.org/10.1016/j.bbi.2023.01.010>. [PubMed]

152. Lei Y, Zhang J, Schiavon CR, He M, Chen L, Shen H, Zhang Y, Yin Q, Cho Y, Andrade L, Shadel GS, Hepokoski M, Lei T, et al. SARS-CoV-2 Spike Protein Impairs Endothelial Function via Downregulation of ACE 2. *Circ Res.* 2021; 128:1323–26. <https://doi.org/10.1161/CIRCRESAHA.121.318902>. [PubMed]

153. Perico L, Morigi M, Galbusera M, Pezzotta A, Gastoldi S, Imberti B, Perna A, Ruggenenti P, Donadelli R, Benigni A, Remuzzi G. SARS-CoV-2 Spike Protein 1 Activates Microvascular Endothelial Cells and Complement System Leading to Platelet Aggregation. *Front Immunol.* 2022; 13:827146. <https://doi.org/10.3389/fimmu.2022.827146>. [PubMed]

154. Perico L, Morigi M, Pezzotta A, Locatelli M, Imberti B, Corna D, Cerullo D, Benigni A, Remuzzi G. SARS-CoV-2 spike protein induces lung endothelial cell dysfunction and thrombo-inflammation depending on the C3a/C3a receptor signalling. *Sci Rep.* 2023; 13:11392. <https://doi.org/10.1038/s41598-023-38382-5>. [PubMed]

155. Robles JP, Zamora M, Adan-Castro E, Siqueiros-Marquez L, Martinez de la Escalera G, Clapp C. The spike protein of SARS-CoV-2 induces endothelial inflammation through integrin $\alpha 5\beta 1$ and NF- κB signaling. *J Biol Chem.* 2022; 298:101695. <https://doi.org/10.1016/j.jbc.2022.101695>. [PubMed]

156. Ghazanfari D, Courreges MC, Belinski LE, Hogrell MJ, Lloyd J, C Bergmeier S, McCall KD, Goetz DJ. Mechanistic insights into SARS-CoV-2 spike protein induction of the

chemokine CXCL10. *Sci Rep.* 2024; 14:11179. <https://doi.org/10.1038/s41598-024-61906-6>. [PubMed]

157. Gultom M, Lin L, Brandt CB, Milusev A, Despont A, Shaw J, Döring Y, Luo Y, Rieben R. Sustained Vascular Inflammatory Effects of SARS-CoV-2 Spike Protein on Human Endothelial Cells. *Inflammation.* 2025; 48:2531–47. <https://doi.org/10.1007/s10753-024-02208-x>. [PubMed]

158. Li H, Wan L, Liu M, Ma E, Huang L, Yang Y, Li Q, Fang Y, Li J, Han B, Zhang C, Sun L, Hou X, et al. SARS-CoV-2 spike-induced syncytia are senescent and contribute to exacerbated heart failure. *PLoS Pathog.* 2024; 20:e1012291. <https://doi.org/10.1371/journal.ppat.1012291>. [PubMed]

159. Jaiswal A, Shrivastav S, Kushwaha HR, Chaturvedi R, Singh RP. Oncogenic potential of SARS-CoV-2-targeting hallmarks of cancer pathways. *Cell Commun Signal.* 2024; 22:447. <https://doi.org/10.1186/s12964-024-01818-0>.

160. Grand RJ. SARS-CoV-2 and the DNA damage response. *J Gen Virol.* 2023; 104:001918. <https://doi.org/10.1099/jgv.0.001918>. [PubMed]

161. Kim MJ, Kim JY, Shin JH, Son J, Kang Y, Jeong SK, Kim DH, Kim KH, Chun E, Lee KY. The SARS-CoV-2 spike protein induces lung cancer migration and invasion in a TLR2-dependent manner. *Cancer Commun (Lond).* 2024; 44:273–77. <https://doi.org/10.1002/cac2.12485>. [PubMed]

162. Moutal A, Martin LF, Boiron L, Gomez K, Ran D, Zhou Y, Stratton HJ, Cai S, Luo S, Gonzalez KB, Perez-Miller S, Patwardhan A, Ibrahim MM, Khanna R. SARS-CoV-2 spike protein co-opts VEGF-A/neuropilin-1 receptor signaling to induce analgesia. *Pain.* 2021; 162:243–52. <https://doi.org/10.1097/j.pain.0000000000002097>. [PubMed]

163. Devaux CA, Camoin-Jau L. Molecular Mimicry of the Viral Spike in the SARS-CoV-2 Vaccine Possibly Triggers Transient Dysregulation of ACE2, Leading to Vascular and Coagulation Dysfunction Similar to SARS-CoV-2 Infection. *Viruses.* 2023; 15:1045. <https://doi.org/10.3390/v15051045>. [PubMed]

164. Zhang S, El-Deiry WS. Transfected SARS-CoV-2 spike DNA for mammalian cell expression inhibits p53 activation of p21(WAF1), TRAIL Death Receptor DR5 and MDM2 proteins in cancer cells and increases cancer cell viability after chemotherapy exposure. *Oncotarget.* 2024; 15:275–84. <https://doi.org/10.18632/oncotarget.28582>. [PubMed]

165. Gioia U, Tavella S, Martínez-Orellana P, Cicio G, Colliva A, Ceccon M, Cabrini M, Henriquez AC, Fumagalli V, Paldino A, Presot E, Rajasekharan S, Iacomino N, et al. SARS-CoV-2 infection induces DNA damage, through CHK1 degradation and impaired 53BP1 recruitment, and cellular senescence. *Nat Cell Biol.* 2023; 25:550–64. <https://doi.org/10.1038/s41556-023-01096-x>. [PubMed]

166. Tsuji S, Minami S, Hashimoto R, Konishi Y, Suzuki T, Kondo T, Sasai M, Torii S, Ono C, Shichinohe S, Sato S, Wakita M, Okumura S, et al. SARS-CoV-2 infection triggers paracrine senescence and leads to a sustained senescence-associated inflammatory response. *Nat Aging.* 2022; 2:115–24. <https://doi.org/10.1038/s43587-022-00170-7>. [PubMed]

167. Ogata AF, Cheng CA, Desjardins M, Senussi Y, Sherman AC, Powell M, Novack L, Von S, Li X, Baden LR, Walt DR. Circulating Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Vaccine Antigen Detected in the Plasma of mRNA-1273 Vaccine Recipients. *Clin Infect Dis.* 2022; 74:715–18. <https://doi.org/10.1093/cid/ciab465>. [PubMed]

168. Fertig TE, Chitoiu L, Marta DS, Ionescu VS, Cismasiu VB, Radu E, Angheluta G, Dobre M, Serbanescu A, Hinescu ME, Gherghiceanu M. Vaccine mRNA Can Be Detected in Blood at 15 Days Post-Vaccination. *Biomedicines.* 2022; 10:1538. <https://doi.org/10.3390/biomedicines10071538>. [PubMed]

169. Yamamoto M, Kase M, Sano H, Kamijima R, Sano S. Persistent varicella zoster virus infection following mRNA COVID-19 vaccination was associated with the presence of encoded spike protein in the lesion. *J Allergy Clin Immunol.* 2022; 6:18–23. <https://doi.org/10.1002/cia2.12278>.

170. Brogna C, Cristoni S, Marino G, Montano L, Viduto V, Fabrowski M, Lettieri G, Piscopo M. Detection of recombinant Spike protein in the blood of individuals vaccinated against SARS-CoV-2: Possible molecular mechanisms. *Proteomics Clin Appl.* 2023; 17:e2300048. <https://doi.org/10.1002/prca.202300048>. [PubMed]

171. Chakraborty D, Singh R, Rajmani RS, Kumar S, Ringe RP, Varadarajan R. Stabilizing Prefusion SARS-CoV-2 Spike by Destabilizing the Postfusion Conformation. *Vaccines (Basel).* 2025; 13:315. <https://doi.org/10.3390/vaccines13030315>. [PubMed]

172. Rong Z, Mai H, Ebert G, Kapoor S, Puelles VG, Czogalla J, Hu S, Su J, Prtvar D, Singh I, Schädler J, Delbridge C, Steinke H, et al. Persistence of spike protein at the skull-meninges-brain axis may contribute to the neurological sequelae of COVID-19. *Cell Host Microbe.* 2024; 32:2112–30.e10. <https://doi.org/10.1016/j.chom.2024.11.007>. [PubMed]

173. Report of the WHO Study Group on Cell Substrates for Production of Biologicals. World Health Organization. 2024.

174. Lenk R, Kleindienst W, Szabó GT, Baiersdörfer M, Boros G, Keller JM, Mahiny AJ, Vlatkovic I. Understanding the impact of *in vitro* transcription byproducts and contaminants. *Front Mol Biosci.* 2024; 11:1426129. <https://doi.org/10.3389/fmbo.2024.1426129>. [PubMed]

175. Maurisse R, De Semir D, Emamekhoo H, Bedayat B, Abdolmohammadi A, Parsi H, Gruenert DC. Comparative transfection of DNA into primary and transformed mammalian cells from different lineages. *BMC Biotechnol.* 2010; 10:9. <https://doi.org/10.1186/1472-6750-10-9>. [PubMed]

176. Patel MN, Tiwari S, Wang Y, O'Neill S, Wu J, Omo-Lamai S, Espy C, Chase LS, Majumdar A, Hoffman E, Shah A, Sárközy A, Katzen J, et al. Enabling non-viral

DNA delivery using lipid nanoparticles co-loaded with endogenous anti-inflammatory lipids. *bioRxiv*. 2024; <https://doi.org/10.1101/2024.06.11.598533>.

177. Wolff JA, Malone RW, Williams P, Chong W, Acsadi G, Jani A, Felgner PL. Direct gene transfer into mouse muscle *in vivo*. *Science*. 1990; 247:1465–68. <https://doi.org/10.1126/science.1690918>. [PubMed]

178. Acsadi G, Jiao SS, Jani A, Duke D, Williams P, Chong W, Wolff JA. Direct gene transfer and expression into rat heart *in vivo*. *New Biol*. 1991; 3:71–81. [PubMed]

179. Wang J, Jiao S, Wolff JA, Knechtel SJ. Gene transfer and expression in rat cardiac transplants. *Transplantation*. 1992; 53:703–5. [PubMed]

180. (Comirnaty, I. P.-. Residual DNA Characterization Report. https://www.ghr.agency/wp-content/uploads/2025/08/comirnaty-residual-dna-characterization-report_en.pdf.

181. Strayer D, Branco F, Zern MA, Yam P, Calarota SA, Nichols CN, Zaia JA, Rossi J, Li H, Parashar B, Ghosh S, Chowdhury JR. Durability of transgene expression and vector integration: recombinant SV40-derived gene therapy vectors. *Mol Ther*. 2002; 6:227–37. <https://doi.org/10.1006/mthe.2002.0657>. [PubMed]

182. Ndeupen S, Qin Z, Jacobsen S, Bouteau A, Estanbouli H, Igayártó BZ. The mRNA-LNP platform's lipid nanoparticle component used in preclinical vaccine studies is highly inflammatory. *iScience*. 2021; 24:103479. <https://doi.org/10.1016/j.isci.2021.103479>. [PubMed]

183. Motwani M, Pesiridis S, Fitzgerald KA. DNA sensing by the cGAS-STING pathway in health and disease. *Nat Rev Genet*. 2019; 20:657–74. <https://doi.org/10.1038/s41576-019-0151-1>. [PubMed]

184. Kwon J, Bakhoun SF. The Cytosolic DNA-Sensing cGAS-STING Pathway in Cancer. *Cancer Discov*. 2020; 10:26–39. <https://doi.org/10.1158/2159-8290.CD-19-0761>. [PubMed]

185. Yang H. Establishing acceptable limits of residual DNA. *PDA J Pharm Sci Technol*. 2013; 67:155–63. <https://doi.org/10.5731/pdajpst.2013.00910>. [PubMed]

186. El-Deiry WS. A pancancer analysis of impact of MDM2/MDM4 on immune checkpoint blockade ICB. *J Clin Oncol*. 2022; 40:2630.

187. Aronoff TE, El-Deiry WS. *MDM2/MDM4* amplification and *CDKN2A* deletion in metastatic melanoma and glioblastoma multiforme may have implications for targeted therapeutics and immunotherapy. *Am J Cancer Res*. 2022; 12:2102–17. [PubMed]

188. El-Deiry WS, Horowitch B, Lou E, Subbiah V, Carneiro BA, Safran H, Gokaslan ZL, Sobol RW, Mani SA, Lawler SE, Chen CC, Wong ET, Kurzrock R, et al. Therapeutic insights for the aggressive subset of high-grade gliomas (HGG) driven by chromosome 1q32 MDM4-containing amplicon and unmethylated MGMT. *J Clin Oncol*. 2024; 42:2080. https://doi.org/10.1200/JCO.2024.42.16_suppl.2080.

189. Huntington KE, Louie AD, Lee CG, Elias JA, Ross EA, El-Deiry WS. Cytokine ranking via mutual information algorithm correlates cytokine profiles with presenting disease severity in patients infected with SARS-CoV-2. *Elife*. 2021; 10:e64958. <https://doi.org/10.7554/elife.64958>. [PubMed]

190. Park H, Nakamura N, Miyamoto S, Sato Y, Kim KS, Kitagawa K, Kobashi Y, Tani Y, Shimazu Y, Zhao T, Nishikawa Y, Omata F, Kawashima M, et al. Longitudinal antibody titers measured after COVID-19 mRNA vaccination can identify individuals at risk for subsequent infection. *Sci Transl Med*. 2025; 17:eadv4214. <https://doi.org/10.1126/scitranslmed.adv4214>. [PubMed]

191. Zhou L, Huntington K, Zhang S, Carlsen L, So EY, Parker C, Sahin I, Safran H, Kamle S, Lee CM, Geun Lee C, A Elias J, S Campbell K, et al. MEK inhibitors reduce cellular expression of ACE2, pERK, pRb while stimulating NK-mediated cytotoxicity and attenuating inflammatory cytokines relevant to SARS-CoV-2 infection. *Oncotarget*. 2020; 11:4201–23. <https://doi.org/10.18632/oncotarget.27799>. [PubMed]

192. Prabhu VV, Morrow S, Rahman Kawakibi A, Zhou L, Ralff M, Ray J, Jhaveri A, Ferrarini I, Lee Y, Parker C, Zhang Y, Borsuk R, Chang WI, et al. ONC201 and imipridones: Anti-cancer compounds with clinical efficacy. *Neoplasia*. 2020; 22:725–44. <https://doi.org/10.1016/j.neo.2020.09.005>. [PubMed]

193. Strandberg J, Louie A, Lee S, Hahn M, Srinivasan P, George A, De La Cruz A, Zhang L, Hernandez Borrero L, Huntington KE, De La Cruz P, Seyhan AA, Koffer PP, et al. TRAIL agonists rescue mice from radiation-induced lung, skin, or esophageal injury. *J Clin Invest*. 2025; 135:e173649. <https://doi.org/10.1172/JCI173649>. [PubMed]

COVID vaccination and post-infection cancer signals: Evaluating patterns and potential biological mechanisms

SUPPLEMENTARY MATERIALS

Supplementary Table 1: Summary of figure permissions and allowed use

References	Figure	Permission	License/Permission basis	Type of Use Permitted	Journal/Date/Publisher
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COVID vaccination and post-infection cancer signals: Evaluating patterns and potential biological mechanisms

Point-by-Point Response to Reviewers

We thank the reviewers for their thoughtful, detailed, and constructive evaluation of our manuscript, and appreciate their recognition of the importance and timeliness of this topic. Below, we provide a point-by-point response and indicate how the manuscript has been revised or clarified accordingly.

We believe these revisions strengthen the manuscript by sharpening its scope, clarifying its intent as a scoping review, while also preserving the central scientific motivation.

Reviewer #1

Kuperwasser and El-Deiry

COVID vaccination and post-infection cancer signals: Evaluating patterns and potential biological mechanisms

This review is a well written comprehensive approach to one of the most critical questions on the horizon bridging SARS-CoV2 infection, and mRNA vaccination across the globe linked to the promotion or initiation of a range of different cancers. It takes a difficult but first attempt at elucidating many of the underlying complexities that now exist and the critical need to address this evolving issue in the world after such a difficult and highly disastrous pandemic.

The authors have done a stringently addressed this question based on current literature, which in itself was difficult due to the lack or searchable indices. I believe it will be important and timely review that will have wide readership in infectious disease and cancer fields. I have some suggestions to help enhance the overall ease of reading as well as its balance.

1. Some mention should be made about the other major neurological, autoimmune and inflammatory consequences due to the mRNA vaccine a bit more and state that this will be focused on cancer. It mentions them but does not really help put them in context. Discuss them early in a paragraph so that the reader have a more complete view of the major problems that are now being dealt with in the medical field due to vaccination.

We agree and have revised the first paragraph of the Introduction to briefly contextualize the manuscript within the broader spectrum of post-vaccination neurological, autoimmune, and inflammatory syndromes that have been reported in the literature, while clearly stating that the present review focuses specifically on oncologic outcomes.

2. Page 7, fix fractions and percentages across the 69 studies with fractions first followed by percentages to be consistent across the statements.

This has been revised.

3. It may be good to examine the differences in vaccination, infection and both with what differences they may entail on page 10 so that it's clear that there are key differences in how individuals may respond to each.

We agree that distinguishing responses to vaccination alone, infection alone, and combined exposure is important and highly relevant. However, the current literature does not provide sufficiently detailed or standardized individual-level data to support a such a comparison. Most published reports are case-based and lack consistent documentation of prior infection status, timing relative to vaccination, cumulative exposure, baseline immune status, or relevant clinical covariates, precluding meaningful stratified analysis.

Accordingly, while we describe vaccination-associated, infection-associated, and combined exposure reports where available, we feel that attempting to infer differential biological responses across these groups is beyond the scope of the paper.

We have a comment in the paper about this limitation.

4. Some mechanisms in HHV-8 may lead to bypass of cell cycle arrest during abortive lytic infection and expression of some lytic antigens including VGPCR (Mesri et al) or Spike induced lytic reactivation.

We thank the reviewer for this important insight.

5. I would suggest to discuss the examples of cancers based on categories in figure 2 with better organization of the figure panels in figure 3 (separate into groups based on cancer types). It's a bit difficult to follow as arranged. The table should be separated out of the figure.

We have reorganized Figure 3 into panels grouped by cancer type (lymphoma, sarcoma, carcinoma, melanoma, glioblastoma and other) based on categories in figure 2. We have also moved the tabular content into a standalone table to improve readability. We have also revised the revised text to parallel the grouping of the cancer types.

6. HIV is a major contributor but I don't believe its accepted as yet that it's a direct causative agent of cancer. Some thinks that way and others still have not accepted this as true.

We have revised this to "HIV is strongly associated with Kaposi's sarcoma, cervical cancer, lymphoma, anal cancer, and other malignancies, largely though immunosuppression and co-infection with oncogenic viruses."

7. HCMV has strong evidence in its association with glioblastoma and breast. See the use of ganciclovir for treatment of glioblastoma in the NEJM paper). The work unproven should be removed and replaced with a more nuanced statement that geographic and molecular methodological differences across studies have hampered consistency across findings.

We revised the wording to reflect available evidence linking HCMV to glioblastoma and breast cancer.

8. Page 173rd paragraph. I would suggest stating that vaccinations were mandated across many countries limiting unvaccinated cohorts. This was basically directly linked to the livelihood of many individuals to take a vaccine.

We appreciate (and agree) with the reviewer's point regarding the impact of vaccine mandates on the availability of unvaccinated comparison groups. This issue is already addressed conceptually in the manuscript through discussion of the limited size and representativeness of unvaccinated cohorts and the resulting constraints on epidemiologic inference. We also agree that widespread vaccination policies influenced cohort composition in many countries, but our intent was to describe this limitation in neutral methodological terms rather than to expand into policy or socioeconomic considerations, which fall outside the scope of the present review. We believe the current framing appropriately captures the relevant study-design implications without detracting from the manuscript's scientific focus.

9. Page 21, great point, which should also focus on connecting with the known prolonged recovery for many individuals after vaccinations. Can this be a cytokine storm that existed in these individuals?

It is indeed plausible that immune dysregulation and sustained inflammatory signaling are potential contributors in some individuals, the current literature does not provide sufficient evidence to conclude that a classical or persistent cytokine storm underlies these prolonged recovery states, nor to directly link such mechanisms to the oncologic observations discussed here.

Given the speculative nature of this hypothesis and the absence of consistent mechanistic or longitudinal data connecting prolonged post-vaccination recovery, cytokine dynamics, and cancer-related outcomes, we chose to limit the discussion to well-described immune perturbations supported by the existing literature. We believe this approach maintains appropriate scientific restraint and keeps the manuscript focused on cancer-related observations and mechanisms directly relevant to the scope of this review.

10. Figure 1, please label the color palates for completeness across the vent diagram. You labeled 6 palates and I could 9 or maybe 10 color palates. Please also complete in the legend.

Addressed

11. Figure 2, reorganize into panels A, B, C, D as well as the color palates and cancer types across the 4 panels. Two top and 2 bottom. Its organized with wasted space as shown now.

Addressed

12. Figure 3, remove table (panel D) into a separate table. Reorganize this into cancer types and different figures. It will be easier to navigate as you reorganize the text to match as well.

We appreciate the reviewer's suggestion to reorganize Figure 3 for navigability. We have done this.

13. Figure 4, Can data be included from other cancers besides the hematologic malignancies? It may make it more complete if its available.

Figure 4 is the data from the US Armed Forces Health Surveillance Division (AFHSD) report. They only looked at NHL.

14. Figure 5, include viral antigens from oncogenic viruses that can drive paracrine and other tumor promoting effects...in the bot I suggest you include VGEF, MMPs, TNF signaling know pathways driving oncogenic activities (place next to the vessel and tumor)

We thank the reviewer for the suggestion. We already indicated $TNF\alpha$ as well as $IL1\beta$ and $IL6$ but have included the text "viral antigens" and "MMPs" in the revised figure.

Reviewer #2

This scoping review by Kuperwasser and Deiry identified 69 peer-reviewed publications (January 2020-October 2025) describing cancers temporally associated with COVID-19 vaccination or infection. The evidence base consisted mainly of 333 case-level reports from 27 countries, along with two large population-based cohort studies and one longitudinal cohort study of U.S. military personnel.

Although the authors frame these findings as hypothesis-generating rather than causal, the review does not adequately account for multiple key confounding factors. In addition, their analysis lacks the denominators and reference comparisons needed to estimate risk or incidence. Hence, the claim that the observed temporal patterns are "difficult to attribute to background incidence alone" is not supported by the data.

We thank Reviewer #2 for their detailed critique and for raising important points regarding epidemiologic interpretation, confounding, and study design. We agree that the available literature does not permit estimation of cancer risk or incidence, nor does it allow adjustment for key confounders using denominators or reference populations.

Importantly, however, this manuscript was not designed to estimate risk or to draw causal inferences. Rather, it is a scoping review intended to systematically assemble, categorize, and contextualize published reports of malignancies temporally associated with COVID-19 vaccination or SARS-CoV-2 infection, and to outline biologically plausible mechanisms that could warrant further investigation. We specifically state in the introduction "The goal of this article is to provide factual information from published literature without bias, and without intent to influence any individual's choices regarding vaccines or risk mitigation." We recognize that case reports and small series are inherently subject to reporting bias and lack appropriate denominators.

In fact, these issues are explicitly addressed in the Limitations section of the manuscript, where we state that the predominance of case reports and small series precludes estimation of incidence or relative risk, that observations are highly susceptible to reporting bias, and that the absence of appropriate control populations limits inference regarding background rates or causality.

Thus, the intent of this scoping review is not to control for confounding or to estimate population-level risk, but to catalog and synthesize published reports for early signal detection and hypothesis generation. Accordingly, all mechanistic interpretations are explicitly described as speculative, and the manuscript states that no causal relationship has been demonstrated. We believe the existing Limitations section accurately and transparently frames these constraints but have added the following sentence to the start of the Results section to avoid any confusion "This scoping review covering the period of January 2020 until April 2025 was not designed to estimate cancer risk or incidence, nor to draw causal inferences, but rather to systematically assemble, categorize, and contextualize published reports of malignancies temporally associated with COVID-19 vaccination or SARS-CoV-2 infection." In addition, we have added the following statement in the conclusion section: "The goal of this review is not to estimate population-level cancer risk but to

provide a structured synthesis of the existing peer-reviewed literature, identify recurring clinical and biological themes, and delineate critical gaps that require rigorous epidemiologic and mechanistic follow-up. This will enable a better understanding of the full spectrum of immune responses to inform safer immunization strategies and illuminate previously underappreciated links between immunity and cancer biology.”

To address this issue (and the subsequent comments below) we have revised the statement in the introduction to state: “The goal of this article is to systematically synthesize and contextualize findings from the published literature regarding malignancies temporally associated with COVID-19 vaccination or SARS-CoV-2 infection, without attempting to estimate risk, establish causality, or inform individual clinical or vaccination decisions.”

Major Comments:

1. Overall evidence base: The Results section is dominated by counts of case reports and small series (333 patients across 69 publications), but there is no corresponding denominator (e.g., number of vaccinated or infected individuals) or comparison to expected background cancer incidence. Without such reference rates, these data cannot support any inference about increased risk.

We agree with this comment, which is why we explicitly state in the conclusion “*The goal is to understand the full spectrum of immune responses to inform safer immunization strategies and illuminate previously underappreciated links between immunity and cancer biology.*” **The goal of this review is not to estimate risk or incidence or provide denominators as this is beyond the scope of the current literature base.** As stated above, we have also revised the language in the introduction to explicitly frame the manuscript as a scoping review for early signal detection and hypothesis generation, consistent with established methodologies and does not interpret the evidence nor make claims as to what they may or may not imply. Nowhere in the manuscript are there claims that vaccination increase risk of cancer. We have strengthened or added language throughout to avoid any implication of risk quantification.

2. Geographic distribution: The authors state that the broad geographic distribution “indicates that the reported temporal associations... are not confined to a particular region or healthcare system.” Given global COVID spread and vaccine rollout, it is expected that cases be reported from multiple countries. Without the proper denominators (e.g., country-level vaccination rates) or standardized incidence comparisons, the geographic spread primarily reflects patterns of case reporting, not evidence of a biological signal.

Again, as stated above, geographic distribution of the published literature simply reflects reporting breadth rather than evidence of differential risk. The intent of this section is to demonstrate that published observations are not confined to a single healthcare system or region.

3. Exposure types: The breakdown of case reports by vaccine platform is purely descriptive. Counts of reports is not the same as risk by platform. The author does not compare the fraction of global doses by platform to the fraction of reported cases.

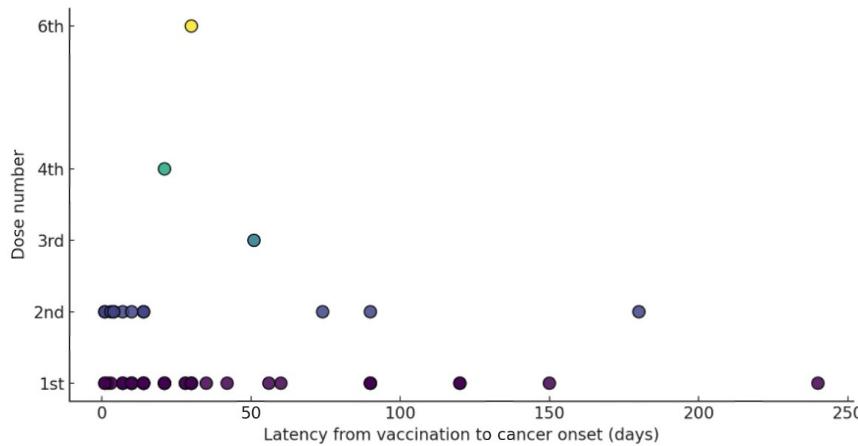
We agree that the vaccine platform breakdown is descriptive only and reflects vaccine availability and uptake rather than comparative risk. We have added the following language to avoid confusion on this point “*This distribution indicates that the published literature is heavily weighted toward mRNA vaccine platforms, particularly Pfizer-BioNTech and Moderna, which together account for the vast majority of vaccine-associated reports. This pattern closely mirrors global vaccination practices where mRNA vaccines were most widely deployed. The relatively smaller representation of adenoviral vector vaccines and inactivated platforms likely reflects both their more limited use in certain regions and differential reporting practices, rather than a comparative assessment of biological risk.*”

4. Cancer types: The statement that, “... making their temporal clustering around vaccination events unusual and difficult to attribute to background incidence alone” is not supported by data. Their argument is based primarily on case reports, which are inherently biased toward unusual reports. Moreover, the authors are counting publications (e.g., 43% of publications reported lymphoid malignancies) rather than patients, and there is no formal analysis of the time from vaccination/infection to diagnosis, nor any comparison with reference distributions or expected numbers of cases in specific post-exposure windows.

We have revised the wording of this to emphasize that observations are reported as unusual by the original authors and are hypothesis-generating, rather than evidence of excess incidence beyond background rates.

5. Timing of onset: The description of latency intervals is purely descriptive and based on a highly selected set of case reports, which are more likely to be chosen when timing is short. The observed clustering of diagnoses within a few weeks of vaccination does not in itself provide evidence of causality.

We agree this section is purely descriptive- but was compiled and plotted from data extracted from all of the case reports that included the information (see data below). Because this was derived from case reports in a qualitative way, nowhere do we state that the latency intervals and timing between vaccination and diagnosis establishes causality. Nowhere in this review do we state or establishes causality of cancer from vaccination.



6. Population-level studies: The two retrospective population-level studies are mentioned, but their actual findings (e.g., effect estimates, cancer types) and extent of confounding control are not summarized. It is difficult to claim “the signal warrants further prospective evaluation” without supporting evidence.

We agree that additional information and synthesis of the population level studies is needed. We have therefore, expanded the summary of population-level studies to describe their design, endpoints, and limitations more explicitly, including confounding control and follow-up duration.

7. For the AFHSD report, the analysis is an ecological time-trend in a specific and highly selected population, and may not be suitable for drawing individual-level inferences about vaccine-related risk. The review highlights an increase in certain NHL subtypes, but does not present absolute counts or rates. It does not address alternative explanations such as changes in diagnostic practices, force composition, or pandemic-related healthcare disruptions.

We agree and do not cite this study to draw individual level inferences about vaccine related risk. This section explicitly states, “*The authors did not analyze or attribute the changes in NHL incidence to vaccination or infection, but the temporal sequence provides an epidemiologic framework for future comparative analyses.*” We agree that these data could be confounded by diagnostic practices and healthcare disruptions during the pandemic and explicitly discuss this limitation in the revised manuscript.

8. Related to the comment above, a 2025 meta-analysis clearly documents widespread interruptions in cancer screening, underscoring the importance of considering this confounder. This could also be true for DoD dataset (figure 4) that the authors highlight. <https://doi.org/10.1038/s43018-024-00880-4>

We agree and acknowledge as a limitation.

9. The authors indicate that most of the oncologic effects were associated with vaccination. Could this be due to the inclusion of REACT19 in their database search?

Yes, the REACT19 database was one of several resources used in our search strategy, and when filtered for “Oncology” it contains approximately 199 references. Many of the references overlap with the peer-reviewed literature identified through PubMed, Scopus, Web of Science, and Google Scholar but could not be found using conventional searching on those platforms.

Thus, the rationale for including REACT19 was not to preferentially capture vaccine-associated outcomes, but rather to address a well-recognized limitation in the discoverability of peer-reviewed publications on SARS-CoV-2 infection, COVID-19 vaccination, and cancer. As noted in the Methods and Limitations sections, much of the relevant literature is not consistently indexed with standard Medical Subject Headings (MeSH) or cross-referenced using conventional

oncology or vaccinology search terms. Consequently, reliance on traditional database queries alone fails to retrieve a substantial portion of the existing peer-reviewed case literature.

We explicitly acknowledge in the manuscript that this reliance on publicly available but incompletely indexed literature imposes inherent limitations, including potential reporting bias, incomplete capture, and challenges in verification. We emphasize that this affects both infection- and vaccination-associated reports and reflects structural limitations of current indexing systems rather than an a priori focus on vaccination. Accordingly, the predominance of vaccination-associated reports in the assembled literature should be interpreted as descriptive of the published record to date, not as evidence of differential risk.

Comments on the Methods

1. Aim and estimand: The inclusion criterion of “temporally associated” malignancy after vaccination or infection is reasonable for collecting case materials, but the methods do not clearly distinguish between (i) describing temporal clustering and clinical patterns, and (ii) estimating causal effects on cancer incidence or progression.

The manuscript does not seek to estimate causal effects on cancer incidence or progression. Its aim is explicitly descriptive and hypothesis-generating, consistent with established scoping-review methodologies. We have clearly stated this at the start of the Results section to avoid any implication of risk quantification and to clearly state that incidence estimation lies beyond the scope of the available literature.

2. Use of AI-generated summaries: The methods section describes a general Google search that returned an AI-generated summary stating CDC and NCI statements about vaccine safety. These AI-generated summaries are not curated scientific databases and can be unreliable. They are more as contextual background rather than as part of the primary evidence base.

The reference to an AI-generated Google summary was included strictly as contextual background to illustrate prevailing public-health messaging and the contrast with the peer-reviewed literature. These summaries were not used as data sources, nor were they incorporated into the evidentiary base of the review. All included findings derive from peer-reviewed publications. We have removed this from the paper.

3. Handling of heterogeneous study designs: The authors list the types of studies included but do not provide an analytic plan that describes: (i) how studies will be stratified by design, (ii) which designs are considered suitable for estimating risk versus providing biological plausibility, and (iii) whether any quantitative synthesis of effect measures is planned, and if so, how heterogeneity will be addressed.

Given the dominance of case reports and small series, no quantitative synthesis or meta-analysis was planned or performed. Study designs are described and categorized to contextualize evidentiary weight, with population-level studies clearly distinguished from case-level reports. Only the former are potentially informative for incidence estimation, and even these are discussed cautiously due to confounding and limited follow-up.

4. Confounding assessment: There is no evaluation of confounding in the included observational studies (e.g., adjustment for age, sex, comorbidities, prior cancer, immunosuppressive treatment, infection status, calendar time).

Formal confounding adjustment was not feasible or appropriate given the nature of the underlying literature and the scope of this review. The lack of adjustment for age, comorbidities, prior cancer history, immunosuppression, infection status, and calendar effects is not relevant and we have described the limitations of this review.

The goal of this review is not to control for confounding or to estimate population-level cancer risk, but to provide a structured synthesis of the existing peer-reviewed literature, identify recurring clinical and biological themes, and delineate critical gaps that require rigorous epidemiologic and mechanistic follow-up. We believe the manuscript’s Methods and Limitations sections accurately and transparently frame these constraints and appropriately limit inference.